



## **Date Palm Pollen Grains as a Potential Manager for Male Sub-fertility: A Clinical Trial**

**Husamuldeen S. M. Saeed<sup>1</sup>, Bashier Osman<sup>2</sup>,  
Tarig Muhammed H. El-Hadiyah<sup>2</sup>, Mona S. Mohamed<sup>3\*</sup>,  
Wadah J. A. Osman<sup>3</sup>, Iman H. Abdoon<sup>1</sup> and Ramzi A. Mothana<sup>4</sup>**

<sup>1</sup>Department of Pharmacology, College of Dentistry, University of Tikrit, Tikrit, Iraq.

<sup>2</sup>Department of Pharmacology, Faculty of Pharmacy, University of Khartoum, Khartoum, Sudan.

<sup>3</sup>Department of Pharmacognosy, Faculty of Pharmacy, University of Khartoum, Khartoum, Sudan.

<sup>4</sup>Department of Pharmacognosy, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia.

### **Authors' contributions**

*This work was carried out in collaboration among all authors. Author HSMS performed the experimental part, statistical analysis and discussed the results. Authors BO and TMHEH designed the study and wrote the protocol. Authors MSM and WJAO wrote the first draft of the manuscript. Authors IHA and RAM managed the literature searches and revised and edited the manuscript. All authors read and approved the final manuscript.*

### **Article Information**

DOI:10.9734/JPRI/2020/v32i630451

Editor(s):

(1) Dr. Mohamed Fathy, Assiut University, Egypt.

Reviewers:

(1) Rana Choudhary, Wockhardt Hospital, India.

(2) Olamide Adewale Adesanya, Kampala International University, Uganda.

Complete Peer review History: <http://www.sdiarticle4.com/review-history/57045>

**Original Research Article**

**Received 02 March 2020**

**Accepted 08 May 2020**

**Published 15 May 2020**

### **ABSTRACT**

Medicinal plants are identified and used throughout human history; it has a great economic value especially in drugs discovery. Date palm pollen (DPP) is used traditionally in Sudan for treating sub-fertile male patients. Male infertility is heterogeneous group of disorders, most of them are idiopathic. This study is aimed to investigate the role of pharmaceutical preparation of DPP in amelioration of male sub fertility with detection of any possible adverse effects on the major body system functions, through blood picture, liver enzymes and kidney function. This study is a single group pretest-posttest experimental prospective comparative self-control. Sub-fertile men with Idiopathic oligoasthenozoospermia or azoospermia were received 500 mg capsules of DPP twice daily for three months after conducting their safety profiles to detect any toxic effects on hematological, hepatological and nephrological functions, Blood samples were taken from the patients for serum level of FSH (for azoospermic patients), FSH and Testosterone (for

\*Corresponding author: E-mail: monacom2005@gmail.com;

oligoasthenozoospermic patients). Finally, Semen sample have been obtained for computerized assisted semen analysis (CASA) report I and II. DPP administration induced significant increase ( $p \leq 0.001$ ) in testosterone level (in oligoasthenozoospermic patients) and FSH level (in azoospermic patients). DPP induced significant changes ( $p \leq 0.001$ ) towards improvement in the total and progressive sperm motility percentages measured in oligoasthenozoospermic patients by CASA dynamic analysis report I and II. The toxicological studies for DPP approved their safety use in human.

**Keywords:** DPP; subfertile male; oligoasthenozoospermic; azoospermic patients.

## 1. INTRODUCTION

Infertility is defined as the failure to conceive after regular unprotected intercourse for two years in the absence of known reproductive pathology [1], reflected by defective spermatogenesis due to pituitary disorders, testicular cancer, germ cell aplasia, varicocele, and environmental factors, or due to defective sperm transport resulting from congenital abnormalities, immunological or neurological factors or could be caused by increased incidence of genetic disorders and apoptosis [2].

Oligoasthenozoospermia is characterized by abnormal semen quality with multiple sperm defects [3], while azoospermia is characterized by absent of sperm in the semen [4,5]. There are a number of drugs that can contribute to infertility, but Idiopathic Oligoasthenozoospermia is the commonest cause of male subfertility [6]. No data exist about infertility in Sudan, the big country in Africa [7]. There are few studies on infertility in sub-Saharan Africa, which tend to lack of research due to economic reasons and, possibly, the psychological denial of the problem [8,9].

Most empiric therapies in male sub fertility are hormonal and based upon the theory that increasing the amount of circulating testosterone and/or FSH will improve testicular function, specifically spermatogenesis [10,11]. Non endocrine treatments aimed to improve the quality of semen have been tested or are currently being evaluated; these treatments are varied and include those possible benefits and those without proved effect. Thyroxine, argenine [12], corticosteroids [13], antibiotics [14], methylxanthines [15] and vitamins (as vitamin B<sub>12</sub>, A, E and C) have all been studied [16-18]. Non pharmacological treatments (Elimination of toxic factors and thermal stress, surgical treatments, assisted reproductive technology) are also used [19].

*Phoenix dactylifera* Linn (family Arecaceae) is native to tropical Asia and Africa; it is good source of protein, amino acids, vitamins, dietary fiber, fatty acids, enzymes, hormones and minerals [20]. It also contains carbohydrates, alkaloids, steroids, flavonoids, vitamins and tannins [21,22]. Dates contain at least six vitamins including small amount of vitamin C, vitamin B1, B2, nicotinic acid (niacin) and vitamin A [23]. Applications and uses of DPP grains in traditional and herbal medicine have been recorded throughout history; it has been used in the treatment of sexual incapacity and weakness in the Arab world [24].

Sudan is one of the Arab countries that cultivate date palm trees and rank fifth in production among Arab date producing countries and third in Africa, date cultivation has been confined to Northern and River Nile States of Sudan [25].

## 2. MATERIALS AND METHODS

### 2.1 Plant Material, Patients and Study Design

Date palm pollen (DPP) was prepared as 500 mg capsules in the research and development department of Amipharma laboratories for drugs manufacturing in Khartoum North with quality assurance approval (Appendix 1). Freshly pods of DPP were obtained from Elgold Locality near Dongola, Northern State, Sudan.

The study is single group pretest-posttest experimental prospective comparative self-control; it has been conducted in andrology clinic in the Reproductive Health Care Centre (RHCC), Khartoum-Sudan, from October 2010 to September 2013. Trial was done on subfertile men with age range from 20-60 years with Idiopathic oligoasthenozoospermia or azoospermia, after isolation of known causes and failure of the conventional reproductive therapeutic measures. Patients received 500 mg capsules of DPP twice daily for three months

after conducting their safety profiles to detect any toxic effects on hematological, hepatological and nephrological functions (complete blood picture, Serum alkaline phosphatase enzyme and Serum creatinine).

Blood samples were taken from the patients for serum level of FSH (azoospermic patients), FSH and Testosterone (oligoasthenozoospermic patients). Finally, Semen sample have been obtained by masturbation into sterile containers after an abstinence period of 3-5 days to be sent for testing in the laboratory. Samples were allowed to liquefy at 37°C for 30 minutes and analysed within 1 h of collection for computerized assisted semen analysis (CASA). Analysis was performed according to the WHO strict criteria for motility detailed patterns (WHO 2010), the system follows WHO CASA for Sperm count, motility (total and progressive: report I) and dynamic parameters: report II. After three months the same samples were taken again for analysis and data collection [26].

## 2.2 Assessment of FSH and Testosterone Changes

The hormonal evaluation of FSH and Testosterone was done by radioimmunoassay using special Bio kit .FSH (N.R= 2.1 -18.6 µIU/ml), while Testosterone (N.R =262-870 ng/dl).The procedure and normal range mentioned by the manufacturing company (CIS bio international) were followed.

## 2.3 Assessment of Immunologic, Hematologic and Biochemical Changes

Serum was analysed for the activities of thealkaline phosphatase (ALP) according to Chemiethod [27]. Serum creatinine concentration was measured by a colorimetric method using commercial kit (Randox Laboratories Ltd., U.K.). Haemoglobin concentration, red blood Cells count, white blood cells count, and platelets count were estimated by (Diogon 60 Cell Computerized Haematology System), as tests for any haematological toxicity.

## 2.4 Statistical Analysis

Paired Student T-test through mean and standard deviation were used to demonstrate the significant differences between pre and post treatment values via determination of P value.

Data were statistically analysed by the software computerized program SPSS Version 16. Data were presented in form of tables [28,29].

## 3. RESULTS

### 3.1 Effect of DPP Treatment on Different Parameters in Oligoasthenozoospermic Patients

FSH and Testosterone levels, CASA dynamic parameter analysis report I and II for per-post treated Oligoasthenozoospermic Patientsare shown in Tables (1, 2, 3 and 4).

### 3.2 Effect of DPP Treatment on Different Parameters in Azoospermic Patients

FSH level and CASA dynamic parameter analysis report I for pre-post treated Azoospermic Patients are shown in Tables 5 and 6.

### 3.3 Effect of DPP Treatment on Major Organ Body Systems in Both Oligoasthenozoospermic and Azoospermic Patients

The pre-post values of serum levels of: alkaline phosphate, creatinine, hemoglobin, RBC, WBC and platelets count for both Oligoasthenozoospermic and Azoospermic Patients are shown in Tables 7, 8, 9 and 10.

## 4. DISCUSSION

As shown in Table 1, The oligoasthenozoospermic patients treated with DPP had pre-treatment FSH mean level of 16.71 µIU/ ml which was increased to 16.90 µIU/ ml in the Post-treatment measurement [FSH normal range (N.R) :2.1-18.6 µIU/ml], this changes were with no statistically significant differences. The mean serum level of FSH among oligoasthenozoospermic patients was within normal range in the pre-treatment values and remains normal in the post treatment values Table 1. The result in the current study is contrary to other studies, which found that, DPP causes an increase in serum FSH level in patients with subfertility [30]. Pre-treatment measurement of FSH in azoospermic patients who received DPP was 28.76 µIU/ ml which changed to 28.93 µIU/ ml in the Post-treatment measurement, the change were with no statistically significant differences (Table 5). The

mean serum level of FSH among azoospermic patients was above normal range in the pre-treatment values and this high serum FSH level in men with azoospermia indicates damaged seminiferous tubule [31,32].

The pre-post treatment values of Testosterone [Testosterone N.R:262-870 ng/dl], in oligoasthenozoospermic patients show an increased level of Testosterone with a statistically significant difference ( $p < 0.001$ ), where pre-treatment measurement of Testosterone 493.26 ng/dl has changed to 692.28 ng/dl in the Post-treatment measurement Table 1. A studies show that DPP increased the plasma level of testosterone significantly [30,33-35], and this comes in agreement with the findings of the current research.

DPP contains small amount of vitamin C [36-38], which maintain pituitary-gonadal axis and

testicular structure within normal and alleviate detrimental effects of stress. However, vit.C supplementation maintains the soundness of pituitary-gonadal axis keeping the hormonal pattern within normal ranges and the testicular structure healthy, activating release of both Follicle Stimulating Hormone and Luteinizing Hormone from the anterior pituitary gland which could be a factor in the elevation of testosterone [39,40].

Sperm's count does not change significantly in oligoasthenozoospermic patients after DPP treatment, sperm's count percentage in pre-treatment measures has decreased from 12.37 to 7.59 million/ml in the post treatment measure Table 2 and this comes in agreement with other study that attributes this to the presence of phytoestrogen, as a steroidal component of DPP, which may have influenced sperm parameters negatively [41], because *Phoenix dactylifera*

**Table 1. The pre-post treatment values of FSH and Testosterone in oligoasthenozoospermic patients after DPP treatment**

Group	Measurement group	Mean ±SD	
		FSH µIU/ ml	Testosterone ng/dl
1	Pre-treatment measurement	16.71± 10.34	493.26±176.77
2	Post-treatment measurement	<sup>N.S</sup> 16.90± 11.03	***692.28±172.64

*n*=21 \*\*\*=  $p < 0.001$  (Significant differences) N.S = not significant

**Table 2. The pre-post treatment values of general semen parameters (sperm count, total motility percentage and progressive motility percentage) measured in oligoasthenozoospermic patients after DPP treatment**

Group	Measurement group	Mean ±SD		
		conc. (Mill/ml)	Total motility (%)	Progressive motility(%)
1	Pre-treatment measurement	12.37± 18.36	25.08± 21.81	16.26± 18.36
2	Post-treatment measurement	7.59± 7.25 <sup>N.S</sup>	***48.48± 23.59	***38.38± 20.36

*n*=21 \*\*\*=  $p < 0.001$  (Significant differences) N.S = not significant

**Table 3. The pre-post treatment details and percentages of sperm motility measured (CASA dynamic parameter analysis report I) in oligoasthenozoospermic patients after DPP treatment**

Group	Measurement group	Mean ±SD				
		PR(A)%Class A:Rapid progressive motility	PR(B)%Class B:Slow or sluggish	NP(C)% Class C:Non progressive motility	IM(D)% Class D: Immotile	PR(A+B)%
1	Pre-treatment measurement	9.15± 13.47	7.08± 12.37	8.84± 10.87	70.13± 26.49	16.26± 18.36
2	Post-treatment measurement	***25.54± 18.78	***12.84± 10.84	<sup>N.S</sup> 10.10± 13.17	***51.52± 23.61	***38.38±20.36

*n*=21 \*\*\*=  $p < 0.001$  (Significant differences) N.S = not significant

**Table 4. The pre-post values of sperm tracks and velocities (CASA dynamic parameter analysis report II) after DPP treatment measured in oligoasthenozoospermic patients**

Group	Measurement group	Mean $\pm$ SD		
		VCL ( $\mu$ m/s) Track velocity	VSL( $\mu$ m/s) Progressive velocity	VAP( $\mu$ m/s) Path velocity
1	Pre-treatment measurement	29.59 $\pm$ 21.96	14.94 $\pm$ 13.86	18.30 $\pm$ 15.11
2	Post-treatment measurement	46.92 $\pm$ 19.47***	***29.09 $\pm$ 14.30	***33.45 $\pm$ 15.01

*n=21 \*\*\*= p < 0.001 (Significant differences) N.S = not significant*

**Table 5. The pre-post treatment values of FSH measured in azoospermic patients after DPP treatment**

Group	Measurement group	Mean $\pm$ SD
		FSH $\mu$ IU/ ml
1	Pre-treatment measurement	28.76 $\pm$ (11.90)
2	Post-treatment measurement	28.93 $\pm$ (10.45)N.S

*n=19 N.S = not significant*

(Date) showed an estrogen like activity with different degrees that may be attributed to the presence of sterols. Estrogen regulates the reabsorption of luminal fluid in the head of the epididymis, and presence of excess amount of such compound lead to disruption of this essential function causes the sperm to enter the epididymis in diluted form and reduces the sperm cell's count [42].

As shown in (Table 2), both total motility and progressive motility percentages of sperm motility measured in oligoasthenozoospermic patients stated by CASA dynamic parameter analysis report I after DPP Administration for three months has increased in a statistically significant value ( $p < 0.001$ ), while (Table 3) shows an increased progressive motility PR type A percentage, progressive motility PR type B percentage and progressive motility combined PR (A+B) percentage with a significant decrease in the percentage of immotile spermatozoa percentage (IM% Class D) after DPP Administration for three months in a statistically significant value ( $p < 0.001$ ).

In the current study, it is found that total motility percentage in pre-treatment measures has increased significantly from 25.08 to 48.48%, while it is a progressive motility percentage from 16.26 to 38.38% in the post treatment values (Table 2). According to the improvement in progressive motility percentages in the post treatment value (Table 3), the partitions of progressive motility stated by CASA dynamic parameter analysis report II, has increased significantly as follows: progressive motility PR

type A percentage has increased from 9.15 to 25.54%, progressive motility PR type B percentage increased from 7.08 to 12.84% and progressive motility combined PR (A+B) percentage has increased from 16.26 to 38.38% after DPP Administration for three months in a statistically significant value ( $P < 0.001$ ).

As is shown in (Table 3), non progressive motility (NP) type C percentage after DPP Administration for three months has increased (from 8.84% to 10.10%) but in non-significant value. This increment in the (NP) type C percentage strengthens the percentages of motility partitions are in a positive way towards improving the motility parameters [43-45]. The immotile spermatozoa percentage (IM% Class D) has dropped significantly from 70.13% in the pretreatment measure to 51.52% in the post treatment (Table 3), which can be regarded as a good sign of general sperm motility improvement.

The improvement in sperm motility function in the current study can be attributed to DPP through the antiapoptotic ability of DPP in the urogenital system especially in testis and prostate [46], DPP reduces the sperm DNA denaturation which triggers apoptosis, and therefore, seems to improve its DNA quality, improve sperm quality, and enhance fertility in the male adult rat. Therefore, it may be useful in solving male sub-fertility problems in humans. Several studies of apoptosis on male infertility establish an association between increased cell death and fertility problems [47]. Signs of apoptosis in mature spermatozoa are found primarily associated with the mitochondrial pathway:

activation of initiator caspase-9, disruption of mitochondrial membrane potential, activation of the main executioner caspase-3 and subsequent cellular collapse [48,49]. Increases of reactive oxygen species in tubules and in seminal plasma and of apoptosis are reputed to affect sperm motility [50,51]. Actually, an increase in apoptotic germ cells in testis of patients suffering severe oligozoospermia has been reported [47]. DPP treatment has ameliorated the histopathological and immunohistochemical changes in atypical prostatic hyperplasia (APH) induced in rats [52]. In these prostate diseases, there is an imbalance between prostate cell growth and apoptosis. Moreover, histopathological examination after DPP treatment revealed increased cellular proliferation and reduced apoptosis in ventral prostate. In the current study, motility sperm parameters have been improved probably through the antioxidant activity of DPP, because antioxidants play an important role in protecting semen from ROS and can improve basic sperm parameters in case of idiopathic oligoasthenozoospermia [53]. DPP contain Vitamins E and C [54,38]. Studies have shown that these vitamins possess antioxidant functions, and their inhibitory effects on ROS accumulation vary from one vitamin to the other [55]. As shown in the current study, one of the causes that lie beyond the improvement in sperm motility with DPP therapy could be the control of presence sperm DNA fragmentation as it contain vitamin E which is beneficial in subfertile men and improve sperm DNA fragmentation index and chromatin integrity by reducing sperm DNA damage [56-59]. Vitamins C and E contents of DPP can significantly increase glutathione content, superoxide dismutase and catalase enzyme activity, while also decreasing lipid peroxidation products, total protein and nitric oxide contents and this can explain the improvement in sperm motility parameters after DPP administration in oligoasthenozoospermic patients [60-62]. The significant improvement in sperm motility parameters by DPP administration in the current research can be explained by the

effect of DPP on sperm tail abnormalities, where DPP causes a decrease in sperms with tail abnormalities numbers which interfere with sperm motility [63]. It is a fact that, the tail of the sperm (the flagellum) - confers motility upon the sperm by having the principal components of sperm motility controllers [64].

DPP treatment Effects on sperm tracks and velocities measured in oligoasthenozoospermic patients stated by CASA dynamic parameter analysis report II:VCL (Track velocity) in pre-treatment measures increased significantly from 29.59 to 46.92  $\mu\text{m/s}$ , while (VSL) Progressive velocity increased significantly from 14.94 to 29.09  $\mu\text{m/s}$ . Finally, average path velocity (VAP) increased significantly from 18.30 to 33.45  $\mu\text{m/s}$  in the post treatment values (Table 4).

The three commonly reported CASA parameters include curvilinear velocity (VCL), average path velocity (VAP) and straight line velocity (VSL). These CASA motility parameters have been modelled and refined mathematically to describe the best motion parameters of each spermatozoon as it travels through a microscopic field [65]. The two important movements of spermatozoa, the vigour movement (circular or whiplash) that could be represented by the VCL and the progression movement can be represented by VSL. VAP represents the general trajectory of the spermatozoa [66,67]. A research found that there is significant positive correlations between sperm velocities and sperm mobility [68] and as the motility parameters improved in our current research DPP administration (Table 2 and 3) in oligoasthenozoospermic patients, so that we can conclude that sperm velocities improvement is a reflection for sperm motility improvement. Both the progressive linear motility (VSL) and the vigorous non-linear non- progressive motion (VCL) are of high importance [43], in our current study both increased and improved by the administration of DPP (Table 4) in oligoasthenozoospermic patients.

**Table 6. The pre-post treatment values of general semen parameters (Sperm count, total motility percentage and progressive motility percentage) measured in azoospermic patients after DPP treatment**

Group	Measurement group	Mean $\pm$ SD		
		Conc. (Mill/ml)	Total motility (%)	Progressive motility(%)
1	Pre-treatment measurement	0.000 $\pm$ 0.000	0.000 $\pm$ 0.0000	0.000 $\pm$ 0.000
2	Post-treatment measurement	.000 $\pm$ 0.0000	0.000 $\pm$ 0.000	0.000 $\pm$ 0.000

n=19

**Table 7. The pre-post values of serum levels of alkaline phosphate and creatinine in oligo asthenozoospermic patients after DPP treatment**

Group	Measurement group	Mean $\pm$ SD	
		Serum Conc. of Alk. Phosph. (IU/L)N.R(43-119)	Serum Conc. of Creatinine (mg/dl)N.R(0.6-1.4)
1	Pre-treatment measurement	80.67 $\pm$ 21.19	.876 $\pm$ .2180
2	Post-treatment measurement	91.67 $\pm$ 17.18 <sup>N.S</sup>	0.852 $\pm$ .081 <sup>N.S</sup>

n=21 N.S = not significant

**Table 8. The pre-post values of hemoglobin concentration, RBC, WBC and platelets count measured in oligoasthenozoospermic patients after DPP treatment**

Group	Measurement group	Mean $\pm$ SD			
		Hemoglobin Conc.(gm/dl) N.R(14 $\pm$ 1)	RBC Count $\times 10^6$ /UI N.R (5Mill.-6 Mill.\dl)	WBC Count $\times 10^3$ /UI N.R(4000-11000)	Platelets Count $\times 10^3$ /UI N.R(150000-400000)
1	Pre-treatment measurement	14.03 $\pm$ 1.187	5.37 $\pm$ .56	5.86 $\pm$ 1.280	220.67 $\pm$ 38.351
2	Post-treatment measurement	14.31 $\pm$ 1.352 <sup>N.S</sup>	5.39 $\pm$ .521 <sup>N.S</sup>	6.22 $\pm$ 1.342 <sup>N.S</sup>	231.57 $\pm$ 52.467 <sup>N.S</sup>

n=21 N.S = not significant

**Table 9. The pre-post values of serum levels of alkaline phosphate and creatinine in azoospermic patients after DPP treatment**

Group	Measurement group	Mean $\pm$ SD	
		Serum Conc. of Alk. Phosph. (IU/L)	Serum Conc. of Creatinine (mg/dl)
1	Pre-treatment measurement	91.89 $\pm$ 12.35	.826 $\pm$ .1790
2	Post-treatment measurement	91.89 $\pm$ 12.35 <sup>N.S</sup>	0.805 $\pm$ (.112) <sup>N.S</sup>

n=19 N.S = not significant

**Table 10. The pre-post values of hemoglobin concentration, RBC, WBC and platelets count measured in azoospermic patients after DPP treatment**

Group	Measurement group	Mean $\pm$ SD			
		Hemoglobin Conc.(gm/dl) N.R(14 $\pm$ 1)	RBC Count $\times 10^6$ /UI N.R (5Mill.-6 Mill.\dl)	WBC Count $\times 10^3$ /UI N.R(4000-11000)	Platelets Count $\times 10^3$ /UI N.R(150000-400000)
1	Pre-treatment measurement	13.35 $\pm$ 0.947	5.38 $\pm$ 0.400	5.73 $\pm$ 1.9	224.58 $\pm$ 62.58
2	Post-treatment measurement	13.69 $\pm$ 1.054 <sup>N.S</sup>	5.12 $\pm$ .351 <sup>N.S</sup>	5.54 $\pm$ 1.46 <sup>N.S</sup>	226.00 $\pm$ 63.06 <sup>N.S</sup>

n=19 N.S = not significant

Azoospermic patients were shown no changes in general semen parameters after the treatment (Tables 6). Normal serum level of ALP is [N.R:43-119 IU/L], in the current study, mean value of ALP in the oligoasthenozoospermic patients in the pre-treatment level changed from 80.67 to 91.67 IU/L in the post-treatment values as shown in Table 7, while in azoospermic patients mean value in the pretreatment level changed from 91.89 IU/L to 91.63 IU/L in the post treatment

values as shown in Table 9, in both of the cases, DPP induced a non - significant differences in serum levels of ALP.

The above results in the current research show the safe use of DPP on liver function and this was in agreement with other studies that showed that, DPP significantly reduced CCl<sub>4</sub>-induced elevation in plasma activities of aspartate aminotransferase (AST), alanine

aminotransferase (ALT), alkaline phosphatase (ALP) enzymes and bilirubin concentration and ameliorated morphological and histological liver damage, suggesting that CCl<sub>4</sub>-induced liver damage in rats can be reversed by treatment with DPP and moreover it can also be used prophylactically as a dynamic liver support [69]. DPP also contains safe and effective novel herbal composition for the prevention of liver disorders in acute and chronic alcoholics [70]. However, it is possible that the recorded DPP content of vitamin C may also play a role in hepatoprotection [71,72].

Oligoasthenozoospermic patients showed a normal mean serum creatinine level in the pre-treatment measures 0.876 mg/dl which was changed to 0.852 mg/dl in the post-treatment as shown in Table 7, while azoospermic patients was showed 0.826 mg/dl in pre-treatment and which was changed to 0.805 mg/dl in the post treatment values as shown in Table 9 (Creatinine NR: 0.6-1.4 mg/dl). In both of the cases, DPP induced non - significant differences in serum levels of creatinine. The above results in the current research show the safe use of DPP on renal function and this was in agreement with other studies that show DPP significantly reduces the increase in plasma creatinine and urea concentrations induced by gentamycin nephrotoxic doses and ameliorated the proximal tubular damage. Antioxidant components in the DPP (e.g. melatonin, vitamin E and ascorbic acid) were suggested to be the basis of the nephroprotection [69], while in another recent study they found that, DPP causes significant decrease in serum creatinine due to the ability of DPP to promote the filtration process and increase the efficacy of the two kidneys [73].

In the oligoasthenozoospermic patients the pre-treatment level of hemoglobin concentration changed from 14.03 to 14.31 g/dl in the post-treatment values as shown in Table 8, while in azoospermic patients the mean value in the pretreatment level changed from 13.35 to 13.69 g/dl in the post treatment values as shown in Table 10 (Normal hemoglobin concentration is 14±1 g/dl).

In this study, all the patients have a normal hemoglobin concentration and DPP induced a non-significant changes in both oligoasthenozoospermic and azoospermic patients in hemoglobin concentration. This result is in contrary to the finding of a recent study which claims that, DPP causes significant increase in hemoglobin concentration [73].

The mean value of RBC count in the oligoasthenozoospermic patients included in the current study changed insignificantly, in the pre-treatment level changed from 5.37 to 5.39×10<sup>6</sup> cell/dl in the post-treatment values as shown in Table 8, while in azoospermic patients the mean value in the pretreatment level changed from 5.38 to 5.12 ×10<sup>6</sup> cell/dl in the post treatment values as shown in table 10 (Normal RBCs count NR: 5-6 ×10<sup>6</sup> cell/dl). In the current study, all the patients have a normal RBC count and DPP induced a non-significant change in both oligoasthenozoospermic and azoospermic patients in RBC count. This result is in agreement to the finding of a recent study which claims that, DPP causes no changes in RBCs count [73].

The mean value of WBC count in the oligoasthenozoospermic patients in the pretreatment level changed from 5.86 to 6.22 ×10<sup>3</sup> cell/dl in the post-treatment values as shown in Table 8, while in azoospermic patients the mean value in the pretreatment level changed from 5.73 to 5.54 ×10<sup>3</sup> cell/dl in the post treatment values as shown in Table 10 (WBC count NR: 4-11×10<sup>3</sup> cell/dl). In the current study, all the patients have a normal WBC count and DPP induced a non-significant changes in both oligoasthenozoospermic and azoospermic patients in WBCs count, this result in agreement to the finding of a recent study which claim that, DPP causes no changes in WBCs count.

The mean level of platelet count in the oligoasthenozoospermic patients in the pre-treatment level was changed from 220.67×10<sup>3</sup> cell/dl to 231.57×10<sup>3</sup> cell/dl in the post-treatment as shown in Table 8, while in azoospermic patients mean value in the pretreatment level changed from 224.58 to 226.00 ×10<sup>3</sup> cell/dl in the post treatment values as shown in Table 10 (platelet count NR: 150-400×10<sup>3</sup> cell/dl), in the current study, mean value of platelet count in both of the cases, DPP induce a non - significant differences in platelet count.

## 5. CONCLUSION

Date palm pollen grains administration induced significant increase in testosterone level (in oligoasthenozoospermic patients) and FSH level (in azoospermic patients). It also induced significant changes towards improvement in the total and progressive sperm motility percentages measured in oligoasthenozoospermic patients by CASA dynamic analysis report I and II. The



toxicological studies for DPP approved their safety use in human.

## CONSENT

Written agreement and consent had been taken from the participated patients.

## ETHICAL APPROVAL

The experiment was done in accordance with the Ethical Clearance, approved by the Committee for Ethics at National medicines and poisons board, Republic of Sudan.

## ACKNOWLEDGEMENTS

The authors extend their appreciation to Researchers Supporting Project number (RSP-2019/119), King Saud University, Riyadh, Saudi Arabia for funding this work. The authors are also grateful to the, Faculty of Pharmacy, University of Khartoum, Sudan.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. WHO. WHO laboratory manual for the examination and processing of human semen. WHO Press. 2010;136-140.
2. DGK, Chemie. Alkaline phosphatase. 1972;10:182-192.
3. Feinstein R. Scientific standard in statistical studies. Science. 1988;242: 1257.
4. Mendenhall W. Introduction to probability and statistics. 3rd. Belmont: Wadsworth Publishing Company Inc. 1971;561.
5. Rabee MR. Effect of zinc sulfate and pollen of date palm on men infertility. Kuwait Med. J. 2006;2:16-22.
6. Meeker JD, Godfrey-Bailey L, Hauser R. Relationships between serum hormone levels and semen quality among men from an infertility clinic. J. Androl. 2007;28:397–406.
7. Guyton AC, Hall JE. Textbook of medical physiology. 11th. Philadelphia: W. B. : Saunders Company. 2006;17:1007-1008.
8. Bahmanpour S, Talaei T, Vojdani Z, Panjehshahin MR, Poostpasand LA, Zareei S, Ghaemini M. Effect of phoenix dactylifera Linn pollen grains on sperm parameters and reproductive system of adult male rats. J. Med. Sci. 2006;31:208-212.
9. Abedi A, Parviz M, Karimian SM, Sadeghipour Rodsari HR. The effect of aqueous extract of phoenix dactylifera pollen grain on sexual behavior of male rats. J of Physiology and Pharmacology Advances. 2012;2:235-242.
10. Kostyuk VA, Potapovich AI, Strigunova EN. Experimental evidence that flavonoid metal complexes may act as mimics of superoxide dismutase. Arch Biochem Biophys. 2004;428:204-208.
11. Ziouti AC, Modafar EL, Fleuriet AS, Boustani EL, Macheix JJ. Phenolic compounds in date palm cultivars sensitive and resistant to Fusarium. Biologia Plantarum J. 1996;38:451-457.
12. Eong YJ, Hong FA, Adel A, Kader S, Alyson E. The flavonoid glycosides and procyanidin composition of Deglet Noor dates (Phoenix dactylifera). J. Agric. Food Chem. 2006;54:2405-2411.
13. Al-Shahib W, Marshal RJ. The fruit of the date palm: It's possible use as the best food for the future. Int. J. Food Sci. Nut. 1993;54:247-259.
14. El-Shaieb A, El-Seady Y, Abd-El-Wahab W. Pathological, hematological and biochemical effects of heat stress on broiler with special reference to protective effect of ascorbic acid supplementation. Egypt. J. Basic. Appl. Physiol. 2009;8:91-112.
15. Sönmez M, Türk G, Yüce A. The effect of ascorbic acid supplementation on sperm quality, lipid peroxidation and testosterone levels of male Wistar rats. Theriogenology J. 2005;63:2063-2072.
16. Hess RA, Zhou Q, Nie R. Estrogens and epididymal function. Reprod Fertil Dev J. 2001;13:273-83.
17. Nagwa M Ammar, Sahar Y Al-Okbi, Doha A Mohamed, Lamia T Abou El-Kassem. Food Sciences and Nutrition, Report and Opinion. Pharmacognosy Department. Cairo: National Research centre J. 2009;3-6.
18. Suarez SS. Hamster sperm motility transformations during development of hyperactivation in vitro and epididymal maturation. Gamete Res. 1988;19:51–65.
19. Suarez X and Dai SS. Hyperactivation enhances mouse sperm capacity for penetrating viscoelastic media. Biol. Reprod J. 1996;46:686–691.


20. Votta TJ, Suarez SS, Stauss CR. Sperm motility hyperactivation facilitates penetration of the hamster zona pellucida. *Biol. Reprod J.* 1995;56:1280–1285.
21. Sciarra A, Mariotti G, Salciccia S, Gomez AA, Monti S, Toscano V, Di Silverio F. Prostate growth and inflammation. *J. Steroid Biochem Mol Biol.* 2008;108:254–260.
22. Lin WW, Lamb DJ, Wheeler TM, Abrams J, Lipshultz LI, Kim ED. Apoptotic frequency is increased in spermatogenic maturation arrest and hypospermatogenic states. *J. of Urology.* 1997;158:1791-1793.
23. Paasch U, Grunewald S, Dathe S, Glander HJ. Mitochondria of human spermatozoa are preferentially susceptible to apoptosis. *Annals of the New York Academy of Sciences.* 2004;1030:403-409.
24. Bejarano I, Lozano GM, Ortiz A, Garcia JF, Paredes SD, Rodriguez AB, Pariente JA. Caspase 3 activation in human spermatozoa in response to hydrogen peroxide and progesterone. *Fertil Steril J.* 2008;90:1340-1347.
25. Kao SH, Chao HT, Liu HW, Liao TL, Wei YH. Sperm mitochondrial DNA depletion in men with asthenospermia. *Fertil Steril J.* 2004;82:66–73.
26. Villegas J, Schultz M, Soto L, Iglesias T, Miska W, Sanchez R. Influence of reactive oxygen species produced by activated leukocytes at the level of apoptosis in mature human spermatozoa. *Fertil Steril J.* 2005;83:635–642.
27. Ahmed AE, Shagufta TM, Hisham AM. Anti-inflammatory and antiproliferative activities of date palm pollen (*Phoenix dactylifera*) on experimentally-induced atypical prostatic hyperplasia in rats. *J Inflamm. London.* 2011;8:40.
28. Senka I K, Bojana P. Review of clinical trials on effects of oral antioxidants on basic semen and other parameters in idiopathic oligoasthenozoospermia. *slovenia. BioMed Research Internationa.* 2014;11.
29. Ziouti AC, Modafar EL, Fleuriet AS, Boustani EL, Macheix JJ. Phenolic compounds in date palm cultivars sensitive and resistant to *Fusarium*. *Biologia Plantarum J.* 1996;38:451-457.
30. Kartikeya M, Ashok A, Rakesh S. Oxidative stress & male infertility. *Indian J Med Res.* 2009;129:357-367.
31. Song B, XJ He, Jiang HH, Peng YW, H Wu, Cao YX. Compound Xuanju Capsule combined with vitamin E improves sperm chromatin integrity. *Zhonghua Nan Ke Xue.* 2012;18:1105–1107.
32. Ménézo YJR, Hazout A, Panteix G. Antioxidants to reduce sperm DNA fragmentation: An unexpected adverse effect. *Reproductive BioMedicine Online.* 2007;14:418–421.
33. Greco E, Romano S, Iacobelli M. *ICSI in cases of sperm DNA damage*: Beneficial effect of oral antioxidant treatment. *Human Reproduction J.* 2005;20:2590–2594.
34. Reco E, Iacobelli M, Rienzi L, Ubaldi F, Ferrero S, Tesarik J. Reduction of the incidence of sperm DNA fragmentation by oral antioxidant treatment. *J. of Andrology.* 2005;26:349–353.
35. Oluseyi CA, Sunday O, Rita AO. Influence of Vitamin C and Vitamin E on testicular zinc content and testicular toxicity in lead exposed albino rats. *Nigeria. BMC Pharmacology and Toxicology.* 2012;13.
36. El-Tohamy MM, El-Nattat WS. Effect of antioxidant on lead-induced oxidative damage and reproductive dysfunction in male rabbits. *J Am Sc.* 2010;6:613-622.
37. Adeniyi TT, Ajayi GO, Akinloye OA. Effect of ascorbic acid and *Allium sativum* on tissue lead level in female *Rattus navigicus*. *Niger J Health Biomed Sci.* 2008;7:38-41.
38. Quill AT, Garbers LD. *Fertilization: Sperm motility activation and chemoattraction.* Daniel M. Hardy. California; Academic Press. 2002;29.
39. Al Kharage E, Rokaya A. Protective effects of *Phoenix dactylifera* (date palm) against Cisplatin- induced genotoxicity. *Food Science and Technology Abstracts.* 1982; 1:148-331.
40. Aida M. Cancel, Danelle Lobdell, Pauline Mendola, Sally D. Perreault. Objective evaluation of hyperactivated motility in rat spermatozoa using computer-assisted sperm analysis. *Hum Reprod J.* 2000; 15:1322-1328.
41. Group, ESHRE Andrology Special Interest. Consensus workshop on advanced diagnostic and rology techniques. *Hum. Reprod J.* 1996;11.
42. Yanagimachi R. *Mammalian fertilization: The physiology of reproduction.* [ed.] E Neill, J.D. In Knobil. New York: Raven Press. 1994;189–317.
43. Laura MK, Denise RH, Ann MD. Correlation of CASA velocity and linearity parameters with sperm mobility phenotype in Turkeys

- from the germplasm and gamete physiology laboratory. USA: Maryland. J Andro. 2000;21:65–71.
44. Al-Qarawi AA, Mousa HM, Ali BEH, Abdel-Rahman H, El-Mougy SA. Protective effect of extracts from date (*Phoenix dactylifera* L.) on the carbon tetrachloride –induced hepatotoxicity in rats. Intern.J Appl. Res. Vet. Med. 2004;2:176-180.
  45. Thornfeldt CR, Pastor EK, Inventors LR, Dykas S, Nipper LLP and Assignee US. Treatment of muco - cutaneous disorders through reversing chronic inflammation and barrier disruption. US Patent J. 2006; 2:33-39.
  46. Saunders, Burtis CA. and Ashwood ER. Tietz's Fundamentals of Clinical Chemistry. Philadelphia. 2001;2:565.
  47. Ahmed MB, Hasona NA, Selemain HA. Protective effects of extract from dates (*Phoenix Dactylifera* L.) and ascorbic acid on thioacetamide-induced hepatotoxicity in rats. Iranian J. of Pharmaceutical Research. 2008;7:193-201.
  48. Effect of date palm (*Phoenix dactylifera*) seeds extracts on hematological, biochemical parameters and some fertility indices in male rats. Sahar HO, Sherif MS. Egypt: s.n. Int. J. of Sci.: Basic and Applied Research. 2011;1-11.
  49. Gurunath S, Pandian Z, Richard A. Anderson, Siladitya Bhattacharya. Defining infertility—a systematic review of prevalence studies. Human Reproduction Update. 2011;17:575–588.
  50. Vaithinathan, Shereen Cynthia D'Cruz, P.P. Mathur. *Apoptosis and Male Infertility*. Pathogenesis of male infertility: Ashok Eds. 2012;520-528.
  51. Onyeka CA, Ashiru OA, Duru FI, Olorunfemi O, Fabunmi OO, Oluwatuyi TS. Semen analysis of 263 sample men from infertility clinic in Western Negeria. West Africa J. of assisted reproduction. 2012;1-16.
  52. Foresta C, Ferlin A, Betella A, Rossato M, Varotto A. Diagnostic and clinical features in Azoospermia. Clin J. Endocr. 1995;43: 537-543.
  53. HW., Daniel. Sperm banking and the cancer patient. Ther Adv Urol J. 2010;2: 19–34.
  54. Hirsh, Anthony. Male subfertility. BMJ. , 2003;327:669.
  55. Elsir A Elussein, Yagoub M Magid, Maha M Omer, Ishag Adam. Clinical patterns and major causes of infertility among Sudanese couples. Trop Doct. J. 2008; 38:243–244.
  56. Olatunji AO, Sule-Odu AO. The pattern of infertility cases at a university hospital. West Afr Med J. 2003;22:205–7.
  57. Larsen U, Masenga G, Mlay J. Infertility in a community and clinic-based sample of couples in Moshi, Northern Tanzania. East Afr Med J. 2006;83:10–17.
  58. Boyle, Karen E. Non surgical treatment of male infertility: Empiric theory. Larry I. Lipshultz, Stuart S. Howards, and Craig S. Niederberger. Infertility in the Male: Cambridge University Press. 2009;439.
  59. ESHER Capri Workshop group. Male infertility and subfertility: Guidelines for management. Hum. Reprod J. 1994;9: 1260-1264.
  60. Jungling ML, Bunge RG. The treatment of spermatogenic arrest with arginine. Fertil Steril J. 1976;27:282–3.
  61. DT, Uehling. Low-dose cortisone for male infertility. Fertil Steril J. 1978;29:220–221.
  62. Hellstrom WJ, Schachter J, Sweet RL, McClure RD. Is there a role for Chlamydia trachomatis and genital mycoplasma in male infertility. Fertil Steril J. 1978;48:337–339.
  63. Wang C, Chan CW, Wong KK, Yeung KK. Comparison of the effectiveness of placebo, clomiphene citrate, mesterolone, pentoxifylline, and testosterone rebound therapy for the treatment of idiopathic oligozoospermia. Fertil Steril J. 1983;40: 358–365.
  64. Dawson EB, Harris WA, Teter MC, Powell LC. Effect of ascorbic acid supplementation on the sperm quality of smokers. Fertil. Steril. J. 1992;58:1034-1039.
  65. Suleiman SA, Ali ME, Zaki ZM. Lipid peroxidation and human sperm motility. Protective role of vitamin E. J. 1996;17: 530-537.
  66. Sandler B, Faragher B. Treatment of oligozoospermia with vitamin B12. Infertility. 1984;7:133-138.
  67. Mahdi Sepidarkish, Reza Omani-Samani, Mohammad Ali Mansournia, Mir Saeed Yekaninejad, Azar Mardi-Mamaghani, Samira Vesali, Roya Hosseini, and Saharnaz Nedjat. The casual effect of lifestyle factors on outcomes of assisted reproductive techniques: A protocol study on Iranian infertile couples. Reprod Health. 2018;15: 210.

68. Alferez MJM, Campos MS. Beneficial effect of pollen, calcium and magnesium in rats with nutritional feropenic anemia. J. Agric. Food Chem. 2000;48:5715-5722.
69. Ziouti AC, Modafar EL, Fleuriet AS, Boustani EL, Macheix JJ. Phenolic compounds in date palm cultivars sensitive and resistant to Fusarium. Biologia Plantarum J. 1996;38: 451-457.
70. Eong YJ, Hong FA, Adel A, Kader S, Alyson E. The flavonoid glycosides and procyanidin composition of Deglet Noor dates (*Phoenix dactylifera*). J. Agric. Food Chem. 2006; 54:2405-2411.
71. Al-Shahib W, Marshal R.J. *The fruit of the date palm: it's possible use as the best food for the future.* Int. J. Food Sci. Nut. 1993; 54: 247-259.
72. El-Desoky GE, Ragab AA, Ismail SA, Kamal AE. *Effect of palm pollen grains (Phoenix dactylifera) on sex hormones, proteins, lipids and liver functions.* J.Agric. Sci. 1995; 20: 4249-4268.
73. AOAD. Arab Agric. *Statistics year book.* 1998; 107:18.

### APPENDIX

#### Quality assurance approval (Appendix 1)



**AMIPHARMA Laboratories Ltd.**  
**Quality Control Department**  
*Sheet for Uniformity of Weight of Capsules.*

**Product Name:** \_\_\_\_\_

**Batch No.:** \_\_\_\_\_ **Q.C. No.:** \_\_\_\_\_

**Manufacturing Date:** \_\_\_\_\_ **Expiry Date:** \_\_\_\_\_

**Pharmacopoeia Requirements:**  
 Not more than two of the individual weights deviate from the average weight by more than 10% when the average of the capsule weight is less than 300mg.  
 Not more than two of the individual weights deviate from the average weight by more than 7% when the average of the capsule weight is 300mg or more.

Capsule No.	Filled Capsule Weight g	Empty Capsule Weight g	Quantity of powder (x) g	(x - $\bar{x}$ )	% Deviat average weight
1	0.6534	0.1091	0.5443	0.0054	1.00
2	0.6461	0.1085	0.5376	-0.0013	-0.24
3	0.6613	0.1118	0.5495	0.0106	1.96
4	0.6479	0.1058	0.5421	0.0032	0.58
5	0.6470	0.1073	0.5397	0.0008	0.14
6	0.6415	0.1051	0.5364	-0.0025	-0.47
7	0.6512	0.1122	0.5390	0.0001	0.01
8	0.6358	0.1089	0.5269	-0.0120	-2.23
9	0.6464	0.1038	0.5426	0.0037	0.68
10	0.6410	0.1029	0.5381	-0.0008	-0.15
11	0.6546	0.1106	0.5440	0.0051	0.94
12	0.6432	0.1094	0.5338	-0.0051	-0.94
13	0.6470	0.1086	0.5384	-0.0005	-0.10
14	0.6506	0.1099	0.5407	0.0018	0.33
15	0.6481	0.1154	0.5327	-0.0062	-1.16
16	0.6494	0.1078	0.5416	0.0027	0.50
17	0.6510	0.1082	0.5428	0.0039	0.72
18	0.6582	0.1113	0.5469	0.0080	1.48
19	0.6425	0.1073	0.5352	-0.0037	-0.68
20	0.6273	0.1011	0.5262	-0.0127	-2.38
SUM	12.9435	2.1650			

Total Weight of filled powder ( $\Sigma x$ ) =  g

Mean filled weight ( $\bar{x}$ ) =  g

Standard Deviation (SD) =  g

Relative Standard Deviation (RSD) =  %

**Applies if average weight is less than 300mg:**  
 Number of individual weights deviating by : more than 10%  more than  
**Applies if average weight is 300mg or more:**  
 Number of individual weights deviating by : more than 7.5%  more than

Test Results Passes / Fails

Date: 15/10/11 Analysts: M. Elmaghrabi Q.C. Manager: \_\_\_\_\_

QC calculation

0014N	+	0.6534 g
0024N	+	0.6273 g
0034N	+	0.6461 g
0044N	+	0.6613 g
0054N	+	0.6479 g
0064N	+	0.6470 g
0074N	+	0.6415 g
0084N	+	0.6512 g
0094N	+	0.6358 g
0104N	+	0.6464 g
0114N	+	0.6410 g
0124N	+	0.6546 g
0134N	+	0.6432 g
0144N	+	0.6470 g
0154N	+	0.6506 g
0164N	+	0.6481 g
0174N	+	0.6494 g
0184N	+	0.6510 g
0194N	+	0.6425 g
0204N	+	0.6425 g
n		20
$\bar{x}$		0.64718 g
s		0.00760 g
sdrel		1.17 %
sk		12.9435 g
skn		0.6273 g
max		0.6613 g
Diff		0.3340 g
0014N	+	0.1091 g
0024N	+	0.1011 g
0034N	+	0.1085 g
0044N	+	0.1118 g
0054N	+	0.1098 g
0064N	+	0.1073 g
0074N	+	0.1051 g
0084N	+	0.1122 g
0094N	+	0.1089 g
0104N	+	0.1038 g
0114N	+	0.1029 g
0124N	+	0.1106 g
0134N	+	0.1094 g
0144N	+	0.1096 g
0154N	+	0.1099 g
0164N	+	0.1154 g
0174N	+	0.1078 g
0184N	+	0.1082 g
0194N	+	0.1113 g
0204N	+	0.1073 g
n		20
$\bar{x}$		0.10825 g
s		0.00338 g
sdrel		3.12 %
sk		2.1650 g
skn		0.1011 g
max		0.1154 g
Diff		0.0143 g

## Agreement and consent from the participated patients (Appendix 2)

In the name of God, The merciful, the compassionate

### Declaration of Agreement For Participation in the Research

I, ..... Declare of my own accord, willingly and voluntarily, and in my legally accepted state, to participate in the research

After it was explained to me; which will be carried out by the researcher ..... Address of the participant/evidence of identity.....

Telephone:.....

And, in my full mental faculties I do agree to:

1. My entry into the abovementioned research.
2. And that I shall take the medicine ..... Dosage ..... number of times ..... And I do agree to take a number of ..... sample(s) of my blood or other requested samples in the appropriate times to the research, and to inform the medical team with any complications or side-effects that might occur to me, and I have no objection for the carryout of the necessary medical and diagnostical examination.

And, may God bear witness of what I say.

Signature of volunteer/for his Guardian:.....

Date:.....

## Ethical Clearance (Appendix 3)



© 2020 Saeed et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:  
The peer review history for this paper can be accessed here:  
<http://www.sdiarticle4.com/review-history/57045>