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Abundance and Diversity of Insects Associated with Citrus Orchards in Two Different Agroecological Zones of Ghana

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Authors' contributions

This work was carried out in collaboration between all authors. Author OFA designed the study. performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Authors CAM and RK managed the analyses of the study. Author KAN managed the literature searches. All authors read and approved the final manuscript.

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ABSTRACT

We investigated the abundance and diversity of entomofauna associated with citrus orchards in two different agroecological zones of Ghana. Malaise traps, flight interception traps, pitfall traps, chemical "knock down" and visual observation were used for data collection. We recorded a total of 20, 285 individual insects belonging to 387 species from 107 families and 13 orders. Although, several species of insects were common to both agroecological zones, some were more specific to an orchard of a particular zone. Diversity indices such as Shannon-Wiener index, Pielou's evenness and Margalef index were higher in the Coastal Savannah zone than the Semi-Deciduous Rainforest zone during both the wet and the dry seasons. *Oecophylla longinoda* Latreille was the most dominant insect species in each agroecological zone, however, they were more abundant in the semi-deciduous rainforest than the Coastal Savannah zone. Our study shows that only 9% of all the 387 insects collected were pests of citrus. This indicates that citrus orchards are potential habitats for insect biodiversity conservation. We therefore recommend that management tactics which have less or no negative effects on natural enemies, pollinators among others but can effectively suppress insect pest populations (such as the use of biological control agents, restriction of herbicides and pesticides) should be adopted. Our study has also provided the first comprehensive inventory of insect species associated with citrus agroecosystems serving as a baseline data for further studies to encourage adoption of economically sound integrated pest management approach for citrus production in Ghana.

Keywords: Insect diversity; abundance; integrated pest management; Oecophylla longinoda; citrus.

1. INTRODUCTION

Insects constitute the most dominant component of terrestrial and freshwater biodiversity in terms of species richness, animal biomass and critical ecological functions [1]. They have invaded every niche, except the oceanic benthic zone [2]. Estimates of species richness of insects have been reported to vary from 2 million to as many as 50 million [3]. Insects are very sensitive to human-mediated disturbances, habitat loss, pollution and climate change, and because of their sensitivity several insect taxa are used as indicators of global change [4]. The majority of insects on earth are important to humans: A few are harmful such as agriculture pests and disease vectors whereas others are beneficial such as decomposers, dispersers, pollinators and natural enemies of pests [5].

Pesticides are used in conventional farming to suppress pest populations below economic threshold levels and research has shown that more arthropod taxa were found in non-sprayed fields together with greater numbers of predators such as Coccinellids than in fields treated with broad-spectrum insecticides and herbicides [6]. Reduction in bumblebees and butterflies has been observed in farms with higher pesticide application than those with no or lower pesticide application [7]. In Ghana, control of major pests of citrus relies on pesticide application [8] and herbicide application is one of the weed management practices adopted by farmers [9]. These herbicides do not only deprive insects of their source of food but also directly kill them.

After citrus establishment, insect species colonize and over time progressively increase in

diversity and abundance. Insects which were previously considered as minor pests are emerging as key pests in many agroecosystems. There is also evidence that honeybees and other pollinators, such as flies, butterflies and major bioindicators such as ants and beetles are in decline across the globe [10]. Anthropogenic climate change threats to insect biodiversity are global. The guickened rate of environmental could deterioration lead to a loss of whole taxonomic groups. In Ghana, [11] reported an annotated list of insects associated with citrus plantations at Kade in the Semi-Deciduous Rainforest zone, however, knowledge of diversity and abundance of insects associated with citrus agroecosystems in different agroecological zones is understood.

To manage natural resources, restore disturbed habitats or conserve valuable species of concern. the biodiversity of insects specific areas or target habitats needs to be However. this assessed. requires comprehensive but efficient inventory of the organisms and perhaps their role in the [12]. This underscores ecosystem critical necessity of this biodiversity study to gain an in-depth knowledge of diversity and abundance of insects associated with citrus agroecosystems to help implement measures that ensure the conservation of biodiversity and the maintenance of agricultural lands to enhance agricultural productivity and sustainability in Ghana.

The aim of this study was to determine the diversity and abundance of entomofauna associated with citrus plantations in two different agroecological zones of Ghana.

2. MATERIALS AND METHODS

2.1 Sampling Zone

We conducted our study in citrus orchards (monoculture) in the Semi-Deciduous Rainforest and Coastal Savannah agroecological zones. In both orchards, weed management was by the use of machetes, and both farms were wholly organic with the citrus variety being Late Valencia. In the Semi-Deciduous Rainforest zone, the research was carried out in a citrus plantation (Cl. 25) of the Forest and Horticultural Crops Research Centre (FOHCREC) Okumaning, Kade in the Kwaebibirem District of the Eastern Region of Ghana. This area experiences an annual bimodal rainfall pattern ranging between 1200-1300 mm, temperature range of 25-38℃ [13]. The coordinates of the site were N 0609.473', W 000 54.550' and E: 552ft. The orchard was established 20 years ago with a triangular planting distance of 6m x 6m. The vegetation in the study site consisted mainly Panicum maximum and Pueraria phaseoloides. In the Coastal Savannah zone, the research was carried out in a farmer's plot at Asuansi Agriculture Research Station in the Abura/Aseibu/Kwamankesse District of the Central Region of Ghana. The area experiences a mean annual rainfall of 980 mm and the rainfall pattern follows the bimodal distribution. Mean monthly temperature is about 26.90℃ [14] and the coordinates of the study site were N 05° 18.654', W 001° 15.667' and E: 363ft. The citrus orchard was established 15 years ago with a triangular planting distance of 7 m x 7 m. The predominant vegetation of the study area consisted mainly of Chromolaena odorata and Panicum maximum.

2.2 Insect Collection

Our study was carried out from September 2013 to March 2014. There were eleven main sampling methods used for the insect collection. Sweep net to Sample the topmost part of the vegetation of the understory to collect vegetation-dwelling insects; Aerial nets were used for flying insects such as dragonflies, butterflies and members belonging to the family Cetoniidae; in each agroecological zone, 10 pitfall traps were randomly set to collect ground crawling insects; 10 coloured pan traps (yellow, blue and orange) were randomly set to collect insects which are attracted to a particular colour; and 5 yellow sticky traps were set up to collect flying

insects. One malaise trap was set in each zone to collect mainly flying nocturnal insects; one flight interception trap was used for both ground crawling insects and flying insects by intercepting their flights. Hand picking was used for collecting specifically slow moving insects and those which play dead when the vegetation is disturbed. Chemical knockdown was used to collect insects in the canopy of the citrus trees: 10 trees were randomly selected during each sampling period and sprayed with CYDIM super (an emulsifiable concentrate containing 36 g cypermethrin and 400 g dimethoate active ingredient/L) at a rate of 120 ml insecticide/ha using a motorized mist blower early in the morning between 6:00-10:00 am. Pieces of vinyl sheets measuring 2.8 m x 8 m were placed beneath the selected trees to collect any insect that fell from the trees during and after spraving.

2.3 Identification of Insects

Insects were preserved in 70% ethanol, sorted, identified to the lowest taxonomic rank possible and counted. Identification of insects was done with reference to collection in the museum of the Department of Animal Biology and Conservation Science (DBCS), University of Ghana, as well as with reference to Gullan and Cranston [15-21]. Dr. Maxwell K. Billah of DABCS helped in the identification of the fruit flies. The voucher specimens were deposited at the African Regional Postgraduate Programme in Insect Science (ARPPIS), West African Sub-Regional Centre.

2.4 Data Analysis

Shannon-Wiener diversity index (H') calculated according to the equation: H'=- $\Sigma(ni/N)\log(ni/N)$, where ni= number of individuals in the ith species and N= total number of individuals sampled. Margalef (d=S-1)/Log (N), is a measure of the number of species present, making some allowance for the number of individuals whiles Pielou's Evenness measures how evenly the individuals are distributed among the different species, both were computed using Past 3.01. To check the completeness of inventory in the orchards, EstimateS (version 9.1.0) was used. The insects were put into two different functional groups; pests and others (pollinators, predators, parasitoids and unknown) based on the authors' long term experience of citrus pests in Ghana.

3. RESULTS

A total of 20,285 individual insects belonging to 387 species from 107 families and 13 orders were collected and identified (Table 1). The total number of entomofauna in the Semi-Deciduous Rain Forest zone was higher than that in the Coastal Savannah zone. 11646 individual insects

belonging to 265 species were recorded in the Semi-Deciduous Rain Forest zone compared to 8639 insects from 246 species in the Coastal Savannah zone. The number of citrus pests form both orchards were 35 representing 9% of the total insects collected during the study period. Fig. 1 shows some insects sampled during the study period.



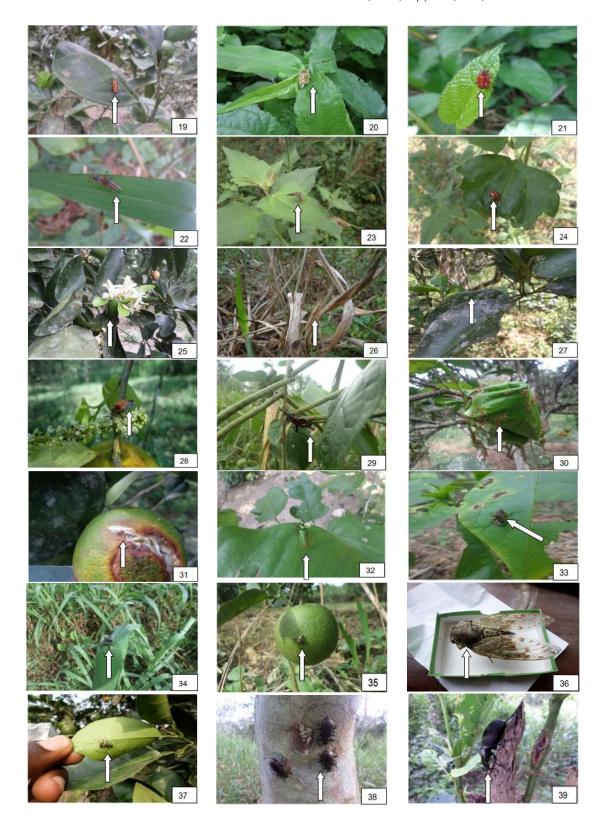




Fig. 1. (1-45) Ophion leteus (1), Brachon sp. (2) Mantis religiosa (3) Sphodromantis sp. (4), Z. variagatus (5), Celaenorrhinus galenus (6), Homoecerus pallens (7), Alydus eurinus (8), Promachus sp. (9), Halyomorpha halys (10), Dragonfly (11), Acraea sp. (12), Green lacewing (13), Zygaena sp. (14), Dysdercus sp. (15) Mylothris sp. (16) Tropidothorax leucopterus (17), Lagria villosa (18), Rhagonycha fulva (19), Lixus sp. (20), Lilioceris lilii (21), Diopsis longicornis (22), Promachus sp. (23), Epilachna bifasciata (24) Marmylida marginella (25), Acrida conica (26), Orophus sp. (27), Lycus trabeatus (28), Sagra femorata (29), Oecophylla longinoda (30) Drosophilla melanogaster (31), Taphronota sp. (32) Laphria sp. (33), Sarcophaga sp. (34), Ceratitis ditissima (35), Cicada sp. (36), Polyrhachis sulcata (37), Coelocnemis lucia (38), Tefflus sp. (39), Leptoglossus sp. (40) Blatella germinica (41), Glymmatophora sp. (42), Cerambycidae sp.(43), Pterophylla camellifolia (44), Madateuchus viettei (45)

Table 1. List of the insect morphospecies and the higher taxonomic ranks of the voucher specimens collected in semi-deciduous rain forest zone (SD) and Coastal Savannah zone (CS) of Ghana

| Order | Family | Morphospecies | SD | CS | Economic status |
|------------|--------------|----------------------------|----|----|-----------------|
| Coleoptera | Scarabaeidae | Madateuchus viettei | 0 | 0 | others |
| | | Scarabaeus viettei Paulian | 1 | 19 | others |
| | | Scarabaeus sp. | 2 | 3 | others |
| | | Caccobius schreberi L. | 0 | 1 | others |
| | | Onthophagus loalien | 0 | 6 | others |
| | | Onthophagus sp. | 2 | 1 | others |
| | | Pseudohammus nyrmedonum | 0 | 5 | others |
| | | Copris sp. | 2 | 9 | others |
| | | Caccobius kelleri | 0 | 3 | others |
| | | Alleucosma viridula Kirby | 0 | 5 | others |
| | Cetoniidae | Pachnoda abyssinica | 0 | 2 | pest |
| | | Marmylida marginella Fab. | 5 | 2 | pest |
| | | Thermophilum fornasini | 0 | 3 | pest |
| | | Torynorrhina flammea | 0 | 3 | pest |
| | | Caelorrrhina barthi | 0 | 1 | pest |
| | | Protaetia fusca | 0 | 3 | pest |
| | | Pachnoda cordata Dury | 0 | 1 | pest |
| | Carabidae | Amara ovara | 1 | 6 | others |
| | | Scaphinotus angusticollis | 0 | 1 | others |
| | | Amara sp. | 2 | 0 | others |
| | | Amara sp 1 | 1 | 0 | others |

| Order | Family | Morphospecies | SD | CS | Economi status |
|-------|-----------------|--------------------------------|----|----|-------------------|
| | | Anomala sp. | 0 | 5 | others |
| | | Tefflus mmegerui | 0 | 2 | others |
| | | Tefflus sp. | 0 | 4 | others |
| | | Harpalus caliginosus Fab. | 1 | 3 | others |
| | | Bembidion patruele Dejean | 3 | 0 | others |
| | | Tetragonoderus deuvei | 4 | 0 | others |
| | Tenebrionidae | Tenebrio sp.1 | 0 | 2 | others |
| | Toriobrioriidae | Alphitobius sp. | 1 | 0 | others |
| | | Tenebrio sp. 2 | 12 | 0 | others |
| | | Coelocnemis sp. | 0 | 1 | others |
| | | | | | |
| | | Coelocnemis lucia Doyen | 1 | 15 | others |
| | | Gonocephalum simplex Fab. | 0 | 10 | others |
| | | Eleodes sp. | 21 | 12 | others |
| | _ | Alobates pennsylvanicus DeGeer | 0 | 1 | others |
| | Chrysomelidae | Chrysochus sp. | 0 | 4 | others |
| | | Lilioceris sp. | 13 | 0 | others |
| | | Lilioceris lilii | 1 | 0 | others |
| | | Ootheca mutabilis | 2 | 6 | others |
| | | Cryptocephalus sp. | 0 | 2 | others |
| | | Podagrica uniformis Jac. | 0 | 7 | others |
| | | Sagara femorata Drury | 5 | 1 | others |
| | | Sagra sp. | 4 | 0 | others |
| | | Cassida sp. | 4 | 1 | others |
| | | • | 4 | 2 | |
| | | Cassida viridus | | | others |
| | | Cassida rubiginosa | 1 | 0 | others |
| | | Nsotra uniformis Jac. | 11 | 0 | others |
| | Cerambycidae | Monochamus sp. | 0 | 7 | pest |
| | | Adynata exilis Borch | 0 | 1 | pest |
| | | Elaphidion sp. | 8 | 0 | others |
| | Coccinelidae | Henosepilachna | 0 | 36 | others |
| | | vigintioctopunctata | | | |
| | | Exochomus sp. | 2 | 20 | others |
| | | Scymnus sp. | 0 | 5 | others |
| | | Chilocorus sp. | 0 | 5 | others |
| | | Cryptocephalus signaticeps | 2 | Ö | others |
| | | Cheilicomus sp. | 2 | Ö | others |
| | | Harmonia axyridis | 1 | Ö | others |
| | | Epilachna bifaciata | 1 | 0 | others |
| | | • | 9 | 0 | _ |
| | O | Epilachna sp. | U | - | others |
| | Curculionidae | Lixus sp. | 0 | 6 | others |
| | | Bryochcta pusilla Pasc. | 3 | 0 | others |
| | | Eloeidobius kamerunicus | 0 | 5 | others |
| | | Sphenophorus maidis Chittenden | 0 | 1 | others |
| | Lagriidae | Lagria hirta | 0 | 1 | others |
| | | Lagria villosa Fab. | 14 | 6 | others |
| | | Cereyonia citri | 2 | 2 | others |
| | | <i>Lagria</i> sp. | 9 | 6 | others |
| | Lycidae | Lycus sp. | 8 | 5 | others |
| | , | Calopteron terminale | 0 | 2 | others |
| | | Lycus trabeatus | 11 | 3 | others |
| | Anobiidae | Stegobium paniceum | 0 | 1 | others |
| | | | | 2 | |
| | Lampyridae | Aspisoma sp. | 0 | | others |
| | Cucujidae | Silvanus oblitus Grouv | 8 | 5 | others |
| | Byrrhidae | Byrrhus fasciatus Forster | 0 | 2 | others |
| | Trogostidae | Phloeobius cordiger Fahrs | 0 | 2 | others |

| Order | Family | Morphospecies | SD | CS | Economic status |
|-------------|-----------------|-----------------------------------|----|-----|-----------------|
| | Cicindelidae | Cicindela leng | 2 | 0 | others |
| | | Cicindella sp. | 6 | 0 | others |
| | Cleridae | Necrobia sp. | 1 | 1 | others |
| | | Necrobia rufibes DeGeer | 0 | 2 | others |
| | Histeridae | Saprinus felipae Lewis | 10 | 0 | others |
| | | Saprinus sp. | 6 | 0 | others |
| | | Spilodiscus biplagiatus Le Conte | 12 | Ö | others |
| | | Platylomalus aequalis Say | 3 | Ö | others |
| | Melolonthidae | Maladera insanabilis Brenske | 10 | 1 | others |
| | Mololofittilaac | Serica sp. | 8 | Ö | others |
| | | Acantthosternum sp. | 0 | 1 | others |
| | Duprostidos | | | | |
| | Buprestidae | Agrilus anxius Gory | 7 | 0 | others |
| | Anthribidae | Araecerus fasciculatus Deg. | 3 | 0 | others |
| | Nitidulidae | Omosita colon Linn. | 7 | 0 | others |
| | Prionidae | Stenodontes sp. | 10 | 0 | others |
| | Elateridae | Cardiophorus sp. | 4 | 0 | others |
| Diptera | Tephritidae | Ceratitis ditissima Munro | 10 | 210 | pest |
| | | Bactrocera invadens | 6 | 11 | pest |
| | | Dacus sp. | 0 | 1 | pest |
| | | Ceratitis cosyra. | 2 | 1 | pest |
| | | Pterandus sp. | 1 | 0 | pest |
| | Therevidae | Evansomyia sp. | 1 | 5 | others |
| | Asilidae | Stenopogon sp. | Ö | 9 | others |
| | Asiliuae | | 8 | 0 | others |
| | | Stichopogon sp. | | | |
| | | Ommatius sp. | 0 | 8 | others |
| | | Laphria sp. | 2 | 0 | others |
| | | Machimus sp. | 2 | 0 | others |
| | | Promachus sp. | 25 | 0 | others |
| | Syrphidae | Eumerus sp. | 0 | 2 | others |
| | | Pseudodorus clavatus Fab. | 0 | 16 | others |
| | | Melannostoma scalare Fab. | 2 | 0 | others |
| | Bibionidae | Bibio albipennis Say | 2 | 1 | others |
| | Drosophilidae | Drosophila melanogaster | 13 | 175 | pest |
| | Diospidae | Diopsis longicornis Macquart | 2 | 4 | others |
| | | Diopsis sp. | 3 | 0 | others |
| | Sarcophagidae | Sarcophaga sp. | 8 | 5 | others |
| | Muscidae | Musca domestica Linn. | 3 | 18 | others |
| | Mascidae | Musca vicina Macq. | 3 | 0 | others |
| | Calinharidaa | • | 2 | 6 | _ |
| | Caliphoridae | Caliphora sp. | _ | - | others |
| | | Lucilia sericata Meigen | 3 | 0 | others |
| | A | Lucilia sp. | 5 | 0 | others |
| | Agromyzidae | <i>Melanagromyza similis</i> Lamb | 3 | 10 | others |
| | Tachinidae | Ramonda spathulata | 4 | 0 | others |
| | Bombyliidae | Bombylius sp. | 3 | 0 | others |
| | Cecidomyiidae | Mayetiola destructor Say | 8 | 0 | others |
| Hymenoptera | Syrphidae | Isodontia mexicana Saussure | 10 | 0 | others |
| - | Mutilidae | Ephutomorpha sp. | 2 | 2 | others |
| | Vespidae | Ėumenes smithii | 4 | 4 | others |
| | • | Eumenes sp. | 4 | 0 | others |
| | | Synagris anali | 2 | 11 | others |
| | | Ropalidia cincta Lepeletier | 7 | 0 | others |
| | | | 0 | 3 | others |
| | | Synagris cornuta | | | |
| | | Polistes sp. | 11 | 0 | others |
| | | Cremnops desertor L. | 3 | 16 | others |
| | | Synagris sp. | 0 | 1 | others |

| Order | Family | Morphospecies | SD | CS | Economic status |
|-----------|---------------|--|-----------|---------|-----------------|
| | Formicidae | Oecophylla longinoda Latri. | 7598 | 4996 | pest |
| | | Odontomachus haematoda Linn. | 54 | 109 | others |
| | | Messor babarus | 1 | 2 | others |
| | | Monomorium Pharaonis Linn. | 45 | 135 | others |
| | | Atopomyrmex crytoceroides Emery | 0 | 85 | pest |
| | | Camponotus pennsylvanicusm | 0 | 15 | pest |
| | | Odontomachus sp. | 46 | 55 | others |
| | | <i>Messor</i> sp. | 4 | 65 | others |
| | | Crematogaster peringueyi Emery Pachycondyla sp. | 1131 9 | 0 11 | pest others |
| | | Crematogaster sp. | 839 | 826 | pest |
| | | Tetramorium sp. | 176 | 232 | pest |
| | | Camponotus sp. | 23 | 94 | others |
| | | Phasmomyrmex aberrans Bolton | 5 | 0 | others |
| | | Polyrhachis sulcata Bolton | 2 | 5 | others |
| | Sphecidae | Ammophila sp. | 2 | 5 | others |
| | • | Amophila insignis Beauv | 8 | 1 | others |
| | | Echilanthus sp. | 1 | 0 | others |
| | | Sphex pensylvanicus Linn. | 2 | 2 | others |
| | Ichneumonidae | Enisoscosphilus sp. | 0 | 5 | others |
| | | Lissonota sp. | 2 | 5 | others |
| | | Echthrus sp. | 0 | 2 | others |
| | | Ophia sp. | 10 | 8 | others |
| | | Anacis sp. | 0 | 2 | others |
| | | Priocnemis conformis Smith | 0 | 1 | others |
| | Apidae | Xylocopa sp. | 22 | 6 | others |
| | | Bombus hortorum L. | 0 | 11 | others |
| | | Xylocopa violacea Linn. | 22 | 6 | others |
| | | Xylocopa virginica Linn. | 3 | 0 | others |
| | Gasterulidae | Gasteruption assectator | 4 | 0 | others |
| | Braconidae | Euphorius sp. | 1 | 0 | others |
| | | Phanerotoma sp. | 0 | 12 | others |
| | | Atanycolus sp. | 0 | 6 | others |
| | | Capitonius sp. | 20 | Ō | others |
| | Chalcididae | Mesocomys pulchriceps Cam | 0 | 4 | others |
| | Evanidae | Evania appendigaster L. | 0 | 4 | others |
| | Melipulidae | Melipona sp. | 1 | 0 | others |
| | Pompilidae | Arachnosphill anceps Gesson | 2 | 0 | others |
| | ' | Trachypompilus sp. | 3 | 0 | others |
| Hemiptera | Coreidae | Leptoglossus oppositus | 0 | 6 | others |
| | | Leptoglossus sp. | 76 | 7 | pest |
| | | Acanthocephala sp. | 0 | 5 | others |
| | | Hypselonotus sp. | 3 | 0 | others |
| | | Homoeocerus pallens F. | 6 | 0 | others |
| | | Homoeocerus sp. | 7 | 0 | others |
| | | Pyrops sp. | 2 | 0 | others |
| | | Leptocorisa acuta Thumb | 2 | 0 | others |
| | Lygaeidae | Spilostethus pandurus | 0 | 2 | others |
| | , , , | Oncopelta fasciatu | Ō | 2 | others |
| | | Oncopelta sp. | 6 | 0 | others |
| | | Mucanum sp. | 4 | Ö | others |
| | | Rhyperochromus sp. | 0 | 2 | others |
| | | Neacoryphus sp. | 3 | 0 | others |
| | | Neacoryphus bicrucis Say | 3 | 0 | others |

| Order | Family | Morphospecies | SD | CS | Economi status |
|-------|---|--|--------|-----|-------------------|
| | | Tropidothorax leucoptrerus Goeze | 0 | 6 | others |
| | Pentatomidae | Oebalus sp. | 0 | 2 | others |
| | | Loxa flavicolis | 0 | 2 | others |
| | | Halyomorpha halys | 6 | 8 | others |
| | | Pentatoma rufipes Linn | 4 | 0 | others |
| | | Euschistus heros | 1 | 4 | others |
| | | Aspavia armigera F. | 1 | 5 | others |
| | | Pentatoma sp. | 4 | 0 | others |
| | | Aspavia albidomaculatus Stall | 5 | 1 | others |
| | | Nezera viridula Linn. | 11 | 0 | others |
| | Saldidae | Pentacora ligata Say | 0 | 15 | others |
| | Reduviidae | Glymmatophora sp. | 21 | 10 | others |
| | Reduvildae | Arilus sp. | 7 | 2 | others |
| | | Saica sp. | 1 | 0 | others |
| | | | | | |
| | | Zelus sp. | 1 | 0 | others |
| | | Rhodius picipes | 2 | 0 | others |
| | | Archilestidium sp. | 0 | 2 | others |
| | | Rapida sp. | 0 | 1 | others |
| | | Tropidothorax leucopterus | 0 | 5 | others |
| | | Reduvius sp. | 0 | 2 | others |
| | | Cydnocoris sp. | 0 | 5 | others |
| | | Nagusta goedeli | 0 | 1 | others |
| | | Stenopoda spinulosa | 4 | 0 | others |
| | | Rhynocoris sp. | 4 | 0 | others |
| | | Triatoma sp. | 6 | 0 | others |
| | Alydidae | Ormenaria rufifascia | 0 | 6 | others |
| | Pyrrhocoridae | Dysdercus superstitious | 9 | 3 | others |
| | ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,, | Cenaeus distinguendus Blote | 0 | 12 | others |
| | | Dysdercus sp. | 6 | 0 | others |
| | Alydidae | Leptocorisa acuta | 0 | 1 | others |
| | riiyalaac | Alydus eurinus Say | 0 | 1 | others |
| | Cercopidae | Neophlaenus sp. | 6 | Ö | others |
| | Cercopidae | Poophilus costalis Walker | 1 | 31 | others |
| | | | | | |
| | | Poophilus sp. | 2 6 | 4 | others |
| | | Neophilaerius sp. | | 0 | others |
| | Marshaadalaa | Plylus grossus L. | 0 | 40 | others |
| | Membracidae | Oxyrachis lamborni Dist. | 0 | 3 | others |
| | | Oxirachis sp. | 1 | 0 | others |
| | | Gargara sp. | 2 | 1 | others |
| | Corixidae | Hesperocorisa sp. | 1 | 0 | others |
| | Aphididae | Aphis gossypii Glover | 32 | 8 | pest |
| | | Toxoptera citricidus Fitch | 19 | 105 | pest |
| | | Aphis maides Fitch | 6 | 0 | pest |
| | | Toxoptera aurantii Boyer | 12 | 88 | pest |
| | Aradidae | Aradius acutus Say | 0 | 10 | others |
| | | Protenor sp. | 0 | 7 | others |
| | Gerridae | Gerris remigis Say | 2 | 0 | others |
| | Issidae | Dieuches armipes Fab. | 19 | 105 | others |
| | Cydnidae | Sehirus cinctus Palisot de Beauvois | 6 | 0 | others |
| | | Pangaeus bilimeatus Say | 4 | 0 | others |
| | Miridae | Brycoropsis laticollis Schum | 0 | 10 | others |
| | | Miridae sp | 1 | 0 | others |
| | | Chamus boxi China | 5 | 0 | others |
| | Psyllidae | Mesohomotoma tessmani (Allum) | 6 | 0 | others |

| Order | Family | Morphospecies | SD | CS | Economic status |
|-------------|----------------|------------------------------------|----|----------------|-----------------|
| | Ricanidae | Ricanidae sp. | 0 | 2 | others |
| | Naucoridae | <i>Ambrysus</i> sp. | 0 | 6 | others |
| | Scuteridae . | Diolcus sp | 12 | 0 | others |
| | Berytidae | Jalysus spinosus Say | 8 | 0 | others |
| | Cicadidae | Cicada orni Boulard | 0 | 6 | others |
| | Dictyopharidae | Taosa sp. | 0 | 3 | others |
| | Coccidae | Lepidosaphes beckii Newn | 45 | 51 | pest |
| | | Planococcus citri Risso | 2 | 6 | pest |
| Lepidoptera | Danaidae | <i>Amauris</i> sp. | 0 | 2 | others |
| | | Danaus chryssipus Linn. | 11 | 5 | others |
| | | Hypolimnas misippus | 2 | 0 | others |
| | | Amauris niavius | 7 | 2 | others |
| | | Amauris albimaculata | 0 | 1 | others |
| | Acraeaidae | Acraea sp. 1 | 4 | 4 | others |
| | 7.0.000.000 | Acraea sp. 2 | 0 | 2 | others |
| | | Acraea pentopolis Ward | Ö | <u>-</u> 16 | others |
| | | Acraea ancedon | 5 | 0 | others |
| | | Acraea perenna | 2 | 0 | others |
| | | Acraea alciope | 7 | 0 | others |
| | Hesperiidae | Calaenorrhinus galenus | 3 | 2 | others |
| | Hespellidae | Pyrrhochelcia iphis | 2 | 0 | others |
| | | | | | others |
| | 7 | Pyrrhochalcia iphis Drury | 0 | 3 | |
| | Zygaenidae | Zygaena sp. | 0 | 3 | others |
| | | Zygaena ephieltes | 2 | 3 | others |
| | | Zygaena sp 1 | 1 | 0 | others |
| | | <i>Zygaena</i> sp 2 | 2 | 0 | others |
| | Noctuidae | Achaea sp. | 98 | 28 | pest |
| | | <i>Achaea obvia</i> Hmps | 0 | 1 | pest |
| | | Anomis leona Schaus | 10 | 3 | others |
| | | Ammalo insulata | 0 | 2 | others |
| | Papilionidae | Papilio demodocus Linn. | 21 | 9 | pest |
| | | Papilio cynorta | 2 | 1 | others |
| | | Papilio nireus | 1 | 1 | others |
| | | Papilio dardanus | 0 | 3 | others |
| | | Papilio conorta | 2 | 1 | others |
| | | Papilio menestheus | 0 | 1 | others |
| | | Pentila pauli Staudinger | 1 | 0 | others |
| | Nymphalidae | Acraea johnstoni | 0 | 1 | others |
| | | Eurytela dryopa Edwards | 3 | 0 | others |
| | | Hypolinmas debius Linn. | 3 | 0 | others |
| | | Hypolimnas misippus Linn. | 7 | Ō | others |
| | | Byblia anvatara R&J | 2 | 13 | others |
| | | Byblia anvatara Boisd | 4 | 0 | others |
| | | Precis pelarga F. | 6 | 2 | others |
| | | Salamis temora | 0 | 1 | others |
| | | Junonia oenone | 14 | 2 | others |
| | | | 0 | 3 | others |
| | | Psuedoacraea sp. Mylanitis leda | 0 | 3 1 | |
| | | Mylanitis leda | | | others |
| | | Bermastistes epaea | 2 | 2 | others |
| | | Achroia sp. | 0 | 1 | others |
| | | Neptis laefa Moore | 3 | 0 | others |
| | | Neptis laeta Oliv. | 0 | 1 | others |
| | | Bicyclus xeneas | 2 | 3 | others |
| | | Erytela sp. | 1 | 0 | others |
| | Arctidae | Arctidae sp. | 0 | 5 | others |

| Order | Family | Morphospecies | SD | CS | Economic status |
|------------|----------------|---|---------|---------|-----------------|
| | | Zale minera | 5 | 0 | others |
| | | Spilosoma mundata Walker | 0 | 5 | others |
| | Pieridae | Dixela doxo | 0 | 1 | others |
| | | Colotis ovagore Klug | 1 | 7 | others |
| | | Eurema briggita Stoll | 9 | 1 | others |
| | | Mylothris poppea Cr. | 1 | 0 | others |
| | | Achiroia sp. | 0 | 1 | others |
| | | Mylothris chloris Fab. | 1 | 3 | others |
| | | Eurema hecabe Linn. | 4 | 3 | others |
| | | Colotis evippe Butler | 2 | 5 | others |
| | Satyridae | Bicyclus safitza Hew | 5 | 2 | others |
| | | <i>Melanis leda</i> Linn. | 1 | 0 | others |
| | | Bicyclus hewitson | 1 | 0 | others |
| | Pyralidae | <i>Pyralia</i> sp. | 2 | 0 | others |
| | • | Homophylotis catori Jord. | 7 | 0 | others |
| | Tinieidae | Ptilobola inornatella Wals | 2 | 0 | others |
| | Gracillariidae | Phyllocnistis citrella Stainto | 0 | 1 | pest |
| | Saturniidae | Eacles imperialis Dury | 0 | 3 | others |
| Orthoptera | Gryllidae | Gryllus lucens Walker | 28 | 24 | others |
| | , | Gryllus pensylvanicus | 0 | 2 | others |
| | | Oecanthus nigricornis | 1 | 5 | others |
| | | Oecanthus sp. | 10 | 1 | others |
| | | Gryllus sp. | 5 | 1 | others |
| | | Gymnogryllus lucens Walker | 6 | 0 | others |
| | Tettigonidae | Atlanticus gibbosus | Ö | 1 | others |
| | rouigomaao | Orophus sp. | 5 | 1 | others |
| | | Hyperhomala woodfordi Kirby | 0 | 5 | others |
| | | Pterophylla camelifora Fab. | 32 | 8 | others |
| | | Conocephalus brevipennis Scudder | 9 | 2 | others |
| | | Tetrix undulata (Sow) | 6 | 3 | others |
| | | Tetrix sp. | 10 | 0 | others |
| | | Tettigonia viridissima Linn | 4 | 0 | others |
| | | Arantia rectifolia | 0 | 8 | others |
| | | Metrioptera roeselii Hagen | 2 | 0 | others |
| | | Conocephalus brevipennis | 0 | 1 | others |
| | | Neoconocephalus ensiger | 0 | 1 | others |
| | | Zabalus lineolatus Stall | 6 | 0 | others |
| | | Scudderia furcata Bruner | 1 | 3 | others |
| | | Mustius seralatus Bolivar | 1 | 0 | others |
| | Acrididae | Neduba carinata | 0 | 1 | others |
| | Autuluae | Froggaltina sp | 0 | 1 | others |
| | | Schistocerca sp. | 6 | 1 | others |
| | | Schistocerca sp. Schistocerca cancellata | 0 | 1 | others |
| | | | 12 | 1 17 | others |
| | | Acrida sp. | | | others |
| | | Acrida conica Melanoplus sp. | 0 5 | 2 | |
| | | | 5 10 | 0 2 | others |
| | | Acanthacris ruficornis Fab. | | | others |
| | | Locusta migratoria | 0 | 3 | pest |
| | | Abisare viridipennis | 3 | 6 | others |
| | | Schistocera albolineata | 11 | 3 | others |
| | | Melanoplus femurrubrum | 2 | 0 | others |
| | | Heteracris littoralis Rambur | 26 | 2 | others |
| | | Gymmobothrus subparallelus | 4 | 0 | others |
| | Pyrgomophidae | Zonocerus variegatus Linn. | 119 | 104 | pest |

| Order | Family | Morphospecies | SD | cs | Economic status |
|-------------|-----------------|--------------------------------------|----|----|-----------------|
| | | Taphronota sp. | 0 | 9 | others |
| | | Attractomorpha similis Bolivar | 7 | 0 | others |
| | | Aractomorpha sp. | 1 | 0 | others |
| | Tetrigidae | Tetrix areuosa Burmeister | 5 | 0 | others |
| | | Pantelia horrenda Walker | 0 | 4 | others |
| | | Tetrix undulate (Sow) | 0 | 4 | others |
| | Catantopidae | Heteracris littoralis Rambur | 3 | 0 | others |
| | | <i>Anacidium</i> sp. | 2 | 0 | others |
| | | Catantops sp. | 13 | 0 | others |
| | Raphidophoridae | Acheta domesticus | 3 | 0 | others |
| | | Ceuthophilus maculatus Harris | 0 | 7 | others |
| | | Centhophillus sp. | 2 | 11 | others |
| Odonata | Libellulidae | Neurothermis sp. | 3 | 2 | others |
| | | Sympetrum corruptum | 2 | 3 | others |
| | | Diplecodes lifebvri Rambur | 2 | 3 | others |
| | | Sympathrum obstrusum (Hagen) | 5 | 0 | others |
| | | Gomphus militaris | 3 | 0 | others |
| | | Plathemis lydia | 1 | 0 | others |
| | | Platheris sp. | 3 | 0 | others |
| | | Libellula quadrimaculata Linn. | 3 | 0 | others |
| | | Erythrodiplax umbrata | 2 | 0 | others |
| | | Brechmorhoga sp. | 4 | 0 | others |
| | Aeshnidae | Anax sp. | 1 | 1 | others |
| | | Anax junius | 3 | 0 | others |
| | Coenogrionidae | Argia apicalis Hagen | 3 | 0 | others |
| | Ü | Agriocnemis fomina fermina Brauer | 6 | 5 | others |
| Dictyoptera | Blattidae | Blatta orientalis (Linn.) | 8 | 14 | others |
| | | Blattella germanica Linn. | 16 | 23 | others |
| | | Blatteria germinica Linn. | 2 | 4 | others |
| | Mantidae | Mantis religiosa Linn | 13 | 12 | others |
| | | Sphodromantis sp. | 17 | 1 | others |
| Isoptera | Termitidae | Macrotermes sp. | 50 | 22 | pest |
| • | | Microtermis sp. | 12 | 23 | pest |
| Psocoptera | Psyllipsocidae | Psyllipscoccus ramburi | 0 | 48 | others |
| Neuroptera | Myrmeleontidae | Brachynemerus abdominalis | 3 | 1 | pest |
| • | • | Brachynemuus sackeni Hagen | 2 | 3 | others |
| Dermaptera | Forficulidae | Forficula auricularia Linn. | 2 | 15 | others |
| • | | Doru aculeatum (Scudder) | 2 | 3 | others |
| Thysanura | Lapismatidae | Lapisma saccharina | 5 | 93 | others |

percentage relative abundance the entomofaunal families collected from citrus orchards in both zones is given in Fig. 2. In the Semi-Deciduous Rain Forest zone, the four most dominant orders were Hymenoptera (86.34%), Hemiptera (3.48%), Orthoptera (3.12%) and Coleoptera (2.34%) whereas Hymenoptera (77.12%), Hemiptera (5.96%), Diptera (5.56%) and Coleoptera (3.15%) were the four most dominant orders in the Coastal Savannah zone (Fig. 1). O. longinoda was the most abundant species recorded in both agroecological zones, recording 7598 (65%) and 4996 (58%) individuals for the Semi-Deciduous Rain Forest and Coastal Savannah zone respectively.

The community indices, specifically Shannon-Wiener (H'), Margalef (d) and Pielou's evenness were calculated for the wet season (September-November) and the dry season (January-March) in each agro-ecological zone (Table 2) as well as the two agro-ecological zones (Table 3). In the wet season, the results show that Margalef

(d=1.341), Shannon-Wiener diversity (H'=1.011) and evenness (0.229) were higher in the Coastal Savannah zone than in the Semi-Deciduous Rain Forest zone. Margalef (d=1.429) and Shannon-Wiener diversity (H' =0.8613) were also higher in the Coastal Savannah zone in the dry season than the Semi-Deciduous-Rain Forest zone. Evenness of species was relatively low for the two agroecological zones in both seasons (Table 2). The results show that all the diversity indices were higher in the Coastal Savannah zone than the Semi-Deciduous Rain Forest zone (Table 3).

Estimated species richness of insects captured using Incidence-based estimator (Chao 1),

Abundance-based estimator (Chao 2), First-order jackknife estimator (Jack 1), Second-order jackknife estimator (Jack 2), Abundance-based coverage Estimator (ACE) and Incidence-based coverage Estimator (ICE) per zone are shown in the species accumulation curves for the Semi-Deciduous Rain Forest zone and Coastal Savannah zone respectively. In both Semi-Deciduous Rain Forest and Coastal Savannah zone, the species accumulation curves were clearly approaching an asymptote showing that species saturation had been reached sampling effort adequate was (Figs. 3 and 4).

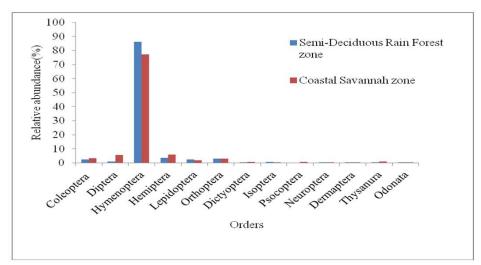


Fig. 2. Relative abundance of insect orders recorded in the study

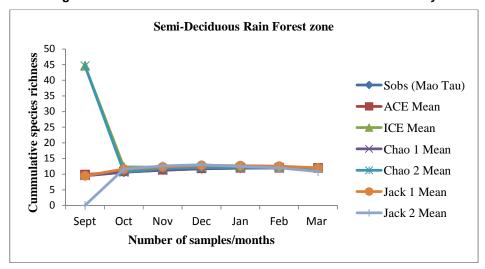


Fig. 3. Species accumulation curve for entomofauna associated with the citrus orchard in the Semi-Deciduous Rain Forest zone and its species richness estimators. Average species richness is based on 50 randomizations [22]

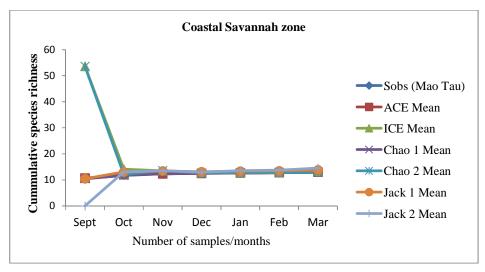


Fig. 4. Species accumulation curve for entomofauna associated with the citrus orchard in the Coastal Savannah zone and its species richness estimators. Average species richness is based on 50 randomizations [22]

Table 2. Diversity indices of the entomofauna for the wet (September-November) and dry (January-March) seasons in the Semi-deciduous rain forest and the Coastal Savannah agroecological zones of Ghana

| Diversity indices | Semi-deciduous rain forest/wet | Coastal Savannah/wet | Semi-deciduous rain forest/dry | Coastal Savannah/dry |
|-------------------|--------------------------------|-------------------------|--------------------------------|-------------------------|
| Shannon, H' | 0.6086 | 1.011 | 0.6309 | 0.8613 |
| Evenness | 0.1671 | 0.229 | 0.1566 | 0.182 |
| Margalef, d | 1.184 | 1.341 | 1.28 | 1.429 |

Table 3. Diversity indices of the entomofauna in the semi-deciduous rain forest and Coastal Savannah agro-ecological zones of Ghana

| Diversity indices | Semi-deciduous rain forest | Coastal savannah | |
|-------------------|----------------------------|---------------------|--|
| Shannon, H' | 0.651 | 0.987 | |
| Evenness | 0.160 | 0.204 | |
| Margalef, d | 1.175 | 1.327 | |

4. DISCUSSION

Our study reports 387 insects compared to 123 insect species that was earlier reported by [12]. The number of insect species found in this present study shows that citrus agroecosystems provide refugium for a plethora of many insect species in Ghana. Our results show that citrus plantations have high diversity and abundance of entomofauna. Most of the insects collected from each agroecological zone were hymenoptera, probably due to the large number of insects belonging to the family formicidae. They were more in the Semi-Deciduous Rainforest zone than the Coastal Savannah zone because the trees in the latter were comparatively younger.

This observation could also probably be attributed to the number of sampling locations, management regimes, sampling techniques and sampling effort in the different studies. This, however, provides the first comprehensive inventory of insects associated with citrus plantations with increasing sampling effort in Ghana.

We found O. longinoda to be the most abundant insect species in each agro-ecological zone. This confirms earlier findings of [23] who reported that Weaver ants O. longinoda tend to be common over most of their ranges. He further noted that in many localities they are among the most abundant and ecologically dominant elements of the arboreal ant fauna. They were, however, more abundant in the semi-deciduous rain forest zone than in the Coastal Savannah zone. probably due to the age differences in the two orchards, citrus trees in the semi-deciduous rain forest zone being older than those in the Coastal Savannah zone. O. longinoda prefers older trees and trees with close canopies [24,25]. Results in the present study are consistent with this earlier

observation. The dominance of O. longinoda over other insect species could also be attributed to the abundance of honeydew-producing insect species such as citrus aphids (Toxoptera aurantii, Toxoptera citricidus, and gossypii), citrus scale insects Lepidosaphes beckii and citrus mealy bugs Planococcus citri [26,27]. The higher number of O. longinoda in the Semi-Deciduous Rainforest zone indicates that citrus trees in this agroecological zone may be better protected from insect pests of citrus than those in the Coastal Savannah zone. It has been earlier noted by [28] that O. longinoda protects crops against insect pests, however, the abundance of O. longinoda could trigger an outbreak of honeydew-producing hemipterans [29] which are not only pests but also transmit plant pathogens [30]. It was also earlier reported that the association between ants and pests such as aphids and scale insects favours the latter by protecting them from their natural enemies [31]. hence could lead to an outbreak of these pests by increasing their abundance and damage.

Our results show that the diversity indices in the Semi-Deciduous Rainforest zone were lower than those in the Coastal Savannah zone. Even though each citrus orchard had several common weeds, some weeds were specific to a particular citrus orchard. The differences could be responsible for variations in abundance and diversity of insect species in each orchard. Variation in the vegetation structure and richness undergrowth indirectly influence entomofaunal abundance and diversity [32]. This also confirms the findings of [33] who earlier reported that the structure of vegetation in different zones may influence the diversity of insects in a particular habitat. Similarly, all the diversity indices were higher in the dry season compared to the wet season. During the wet season, from September-November there were no matured citrus fruits, therefore there were few insects in the orchards but during the dry season, from January to March, the fruits were matured and ripened, hence the higher number of insects in the orchards. This could account for the higher diversity in the dry season than the wet season.

In the Semi-deciduous Rain Forest zone, we observed that the abundance of predatory insects belonging to Formicidae (*Crematogaster* sp., *O. longinoda, Tetramorium* sp.) and Asilidae (*Promachus* sp., *Laphria* sp., *Stichnopogon* sp, *Machimus* sp.,) was more than those in the Coastal Savannah zone. This could also account for the low diversity of insects in the Semi-

Deciduous Rain Forest zone. This had earlier been reported in several studies where the abundance of predators, parasitoids, and the prevalence of diseases are all involved in insect population control [34,35]. We observed that most of the insects associated with citrus orchards in Ghana are not pest. The majority of the insects found in our study were categorized under "others" (predators, pollinators, parasitoids and unknown species).

5. CONCLUSION

The study has provided a species list of insects in citrus plantations in Ghana. The study shows that citrus orchards have diverse and numerous entomofauna with a small proportion being insect pests of citrus. Our study presents the first comprehensive inventory of insects associated with citrus orchards. This may provide a useful foundation for exploring the integrated production and pest management for citrus production. We recommend that to conserve insect species in citrus orchards, management tactics which have less or no negative effects on natural enemies, pollinators and other insects but can effectively suppress insect pest populations (use of biological control agents, restriction of herbicides and pesticides use) should be adopted.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

 Samways MJ. Insect divers. cons. Cambridge: Cambridge University Press. 2005.

- Gramaldi D, Engel MS. Evolution of the insects. Cambridge University Press, New York. 2005;755.
- 3. Stork NE. How many species are there? Biodiversity and Conservation 2. 1993;215-232.
- 4. Menéndez R. How are insects responding to global warming? Tijdschrift voor Entomologie. 2007;150:355-365.
- Raina SK, Chauhan TPS, Tayal MK, Pandey RK, Mohan R. Multiple silkworm cocoon cropping in Jammu division. In Proceedings of the Workshop on Recent Trends in Development of Sericulture in Jammu and Kashmir. 2011;63–67.
- Amalin DM, Peña JE, Duncan R, Leavengood J, Koptur S. Effects of pesticides on the arthropod community in the agricultural areas near the Everglades National Park. Proceedings of the Florida State Horticultural Society 122; 2009.
- 7. Brittain CA, Vighi M, Bommarco R, Settele J, Potts SG. Impacts of a pesticide on pollinator species richness at different spatial scales. Basic and Applied Ecology. 2010;11(2):106-115.
- 8. Afreh-Nuamah K. Pests of citrus speciesrutaceae. In: Obeng-Ofori D. (edited), Major pests of food and selected fruit and industrial crops in West Africa, The City Publishers Ltd. 2007;119-126.
- Ministry of Food and Agriculture. Citrus production in Ghana. Horticulture Exports Industry Initiative (HEII). Crop Services Directorate, Ministry of Food and Agriculture. Agriculture information support unit press. Accra, Ghana. 2007;108.
- 10. Winfree R, Bartomeus I, Cariveau DP. Native pollinators in anthropogenic habitats. Annual review of ecology. Evolution and Systematics. 2011;42:1-22.
- Afreh-Nuamah K. An annotated list of insects associated with citrus plantations at Kade, Ghana. Ghana Journal of Agriculture Science. 1988;24-27.
- 12. Kim KC, Byrne LB. Biodiversity loss and the taxonomic bottleneck: Emerging biodiversity science. Ecological Research. 2006;21:794–810.
- Ofosu-Budu KG. Performance of citrus rootstocks in the forest zone of Ghana. Ghana Journal of Horticulture. 2003;3:1-9.
- 14. Ministry of Food and Agriculture; 2013. Available: www.mofa.gov.gh/site/?page_id= 1436 (Last accessed 30th May, 2016)

- Gullan PJ, Cranston PS. The insects: An outline of entomology. Blackwell, Malden, Mass, USA: 2005.
- Gullan PJ, Cranston PS. Insects: An outline of entomology. Chapman and Hall; 2010.
- Scholtz CH, Holm E. Insects of Southern Africa, Butterworths Professional Publishers: 1989.
- Crowson RA. Coleoptera: Introduction and Key to Families. HIBI. 1956;4.
- Ross HH. A textbook of Entomology. Topan Company Ltd. Tokyo. Japan. 1965.
- McGavin G.C. Insects, Spiders and other Terrestrial Arthropods, Dorling Kindersley. 2002;255.
- 21. Aidoo OF, Kyerematen R, Akotsen-Mensah C, Afreh-Nuamah K. Effect of some climatic factors on insects associated with citrus agro-ecosystems in Ghana. Journal of Biodiversity and Environmental Sciences. 2014;5(4):428-436.
- 22. Colwell RK. Estimates- statistical estimation of species richness and shared species from samples. Version 6. User's guide and application; 2000.

 Available: http://viceroy.eeb.uconn.edu/Estimates
- Magurran, AE. Ecological diversity and its measurement. New Jersey: Princeton University. 1988;197.
- 24. Dejean A, Corbara B, Orivel J, Leponce M. Rainforest canopy ants: The implications of territoriality and predatory behaviour. Function Ecosystem Community. 2007:1:105-120.
- 25. Bawa AS, Yawson GK, Ofori SE, Appiah SO, Afreh-Nuamah K. Relative abundance and diversity of insect species in oil –palm cocoa intercrop at Kusi in the Eastern Region of Ghana. Agriculture Science Research Journal. 2011;1(10):238-247.
- Blüthgen N, Stork NE, Fiedler K. Bottomup control and co-occurrence in complex communities: Honeydew and nectar determine a rainforest ant mosaic. Oikos, 2004;106:344-358.
 - DOI: 10.1111/j.0030-1299.2004.12687.x 2015.01.10
- Tsuji K, Hasyim A, Harlion Nakamura K. Asian weaver ants, Oecophylla smaragdina and their repelling of pollinators. Ecological Research. 2004;19: 669-673.
- 28. Dlussky GM, Wappler T, Wedmann S. New middle Eocene formicid species from

- Germany and the evolution of weaver ants. Acta Palaeontologica Polonica. 2008;53(4):615-626. Available: www.bioone.org/doi/pdf/10.4202/app.2008.04.06
- Holway DA, Lach L, Suarez VA, Tsutsui ND, Case TJ. The causes and consequences of ant invasions. Annual Review of Ecology & System. 2002;33: 181–233.
- Delabie JHC. Trophobiosis between formicidae and hemiptera (Sternorrhyncha and auchenorrhyncha): An overview. Neotropical Entomology. 2001;30:501-515.
- 31. Hill M, Holm K, Vel T, Shah NJ, Matyot P. Impact of the introduced yellow crazy ant *Anoplolepis gracilipes* on Bird Island, Seychelles. Biodiversity and Conservation 2003;12:1969–1984.

- Khadijah AR, Azidah AA, Meor SR. Diversity and abundance of insect species at Kota Damansara Community Forest Reserve, Selangor. Scientific Research & Essays. 2013;8(9):359-374. DOI:10.5897/SRE12.481.2015.01.01
- 33. Abdullah F, Sina I. Rove beetles (Coleoptera: Staphylinidae) of Lanjak Entimau, Sarawak, East Malaysia. International Journal of Zoological Research. 2009;5(3):126-135.
- 34. Koul O, Dhaliwal GS. (eds). Predators and parasitoids. Taylor and Francis, London. (2003);191
- 35. Pinheiro F, Diniz IR, Coelho D, Bandeira MPS. Seasonal pattern of insect abundance in the Brizillian Cerrod. Australia Ecology. 2002;27:132-136.

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