

*International Journal of Plant & Soil Science 8(3): 1-7, 2015; Article no.IJPSS.20270 ISSN: 2320-7035*



**SCIENCEDOMAIN** *international www.sciencedomain.org*

# **Salt Sensitivity in** *Camelina sativa* **Seedlings and Polyamine Content**

# **Roberto Russo<sup>1</sup> and Remo Reggiani<sup>1\*</sup>**

*1 Istituto di Biologia e Biotecnologia Agraria, CNR, Milano, Italy.*

# *Authors' contributions*

*This work was carried out in collaboration between both authors. The authors of this manuscript worked together to design, conduct, analyze and interpret the findings of this experiment. Both authors read and approved the final manuscript.*

#### *Article Information*

DOI: 10.9734/IJPSS/2015/20270 *Editor(s):* (1) Sławomir Borek, Faculty of Biology, Department of Plant Physiology, Adam Mickiewicz University, Poland. *Reviewers:* (1) Edward Missanjo, University of Malawi, Malawi. (2) Rajaram Pandurang Dhok, Savitribai Phule Pune University, Pune, India. Complete Peer review History: http://sciencedomain.org/review-history/10695

*Original Research Article*

*Received 20th July 2015 Accepted 2nd August 2015 Published 24th August 2015*

# **ABSTRACT**

The objective of the present investigation was to study the effect of different NaCl concentrations (0, 50, 75, 100, 125 and 150 mM) on seedlings of *Camelina sativa* (L.) Crantz. On these seedlings, we determined their appearance after 3 days of salt stress, soluble sugar and total amino acid contents and levels of polyamines. The *Camelina* seedlings suffered already at 75 mM of salt and presented more damages at higher concentrations. The content of sugars and amino acids increased up to 75 mM NaCl and then declined. The decrease was especially pronounced on soluble sugars at the highest concentrations. The Polyamine content increased at 50 mM and then decreased at higher concentration. The polyamine increase at 50 mM was mainly due to spermine. The presence of exogenous spermine (1 mM) during the stress ameliorated the tolerance of salinity, confirming previous studies about the protective role of spermine against salt stress.

*Keywords: Camelina sativa; salinity; polyamines; spermine; spermidine; putrescine; soluble sugars; amino acids.*

\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

*\*Corresponding author: E-mail: reggiani@ibba.cnr.it;*

#### **1. INTRODUCTION**

Salinity is the main stress that limits agricultural production. In fact, more than 20% of the cultivated area suffers from salinity and, this percentage, is increasing steadily [1]. The salt stress causes changes in various biochemical and physiological processes depending on the severity of the stress [2,3]. Salinity affects plants through both an osmotic effect (retention of water in the soil) that for a specific effect on the ion regulation [4]. Accumulation of amino acids (mainly proline) and carbohydrates play a role in mitigating the osmotic stress [5-7]. The accumulation in tissues of  $Na<sup>+</sup>$  and Cl<sup>-</sup> is the main detrimental effect in plants exposed to high concentrations of salt [1,8]. These ions cause severe ion imbalance and physiological disorder [1].

The polyamines (PAs) have been suggested to have a specific role in maintaining the cationanion balance in plant cell [8]. Since they are low molecular weight polycations play an important role in ion balancing, especially when CI uptake exceeds the  $Na<sup>+</sup>$  uptake, a frequent condition under salt stress [9,10]. A countermeasure adopted by plants to resist the presence of salt is to modulate the metabolism of PAs although its significance has not yet been well elucidated [11]. Putrescine (Put), spermidine (Spd) and spermine (Spm) are the most abundant PAs in higher plants [12]. In *Arabidopsis thaliana* (species very close to *Camelina*), a mutant lacking Spm caused hypersensitivity to NaCl, possibly due to impaired  $Ca<sup>2+</sup>$ -homeostasis [13]. This phenotype was mitigated by exogenous applied spermine [14].

This study was done to test the sensitivity to salinity of *Camelina sativa* seedlings by assessing their appearance, levels of endogenous solutes (soluble sugars, amino acids) and PA levels.

#### **2. MATERIALS AND METHODS**

Seeds of *Camelina sativa* (L.) Crantz var. Calena were surface sterilized for 5 min with 5% (v/v) NaClO. The seeds were then spread in petri dishes (50 seeds / 10 cm diameter petri) in sterile water for germination. Seven-day-old seedlings were treated for 1 or 3 day with different salt concentrations (0, 50, 75, 100, 125 and 150 mM NaCl). The seedlings from 1 day treatment were then ground in a mortar with: a) 80% (v/v) ethanol at 50°C for 3 hours for amino acids and

soluble sugars analyses; b) 0.6 M PCA (0.1 g fresh weight [FW]/ml PCA) for PAs analysis. The homogenates were cleared by centrifugation at 15000xg for 15 min.

# **2.1 Determination of Total Amino Acids and Soluble Sugars**

The level of total amino acids was determined in alcoholic-extracts by the ninhydrin method according to Moore [15] and further modified by Aurisano et al*.* [16]. Quantitative determination of soluble sugars was with the anthrone method [17]. To 20 µL of alcoholic-extract were added 80 μL of distilled water and 500 μL of anthrone reagent (2 g  $L^{-1}$  of sulfuric acid 72%). Calibration was made with different concentrations of glucose (0, 2, 4, 6, 8 and 10 μg). Samples and standards were heated for 15 min at 100°C in water bath and then cooled rapidly to 0°C on ice. Samples were read at 630 nm within an hour in a plate reader Infinite 200 Pro (Tecan Italia, Cernusco sul Naviglio, Italy).

#### **2.2 Determination of Polyamines**

To 100 μL of PCA extract were added 100 μL of saturated  $Na<sub>2</sub>CO<sub>3</sub>$  and 200 μL of dansyl chloride (5 mg/mL acetone). The reactions in closed tubes were stored overnight in the dark at room temperature. The dansylated PAs were extracted into 200 μL of toluene by vortex mixing. When the two phases were well separated, 20 μL of the toluene layer was loaded with a Linomat IV (Camag, Muttenz, Switzerland) on a HPTLC plate of silica gel 60 with concentration zone (Merck, Darmstadt, Germany). The plate was run using hexane/ethylacetate (11:8, v/v) as solvent. On removal from the tank, the plate was dried for 10 min at 50°C and the fluorescence (excitation 350 nm) of each line read into a densitometer TLC Scanner II (Camag, Muttenz, Switzerland). PAs were identified and quantitated as previously reported [18].

#### **3. RESULTS AND DISCUSSION**

#### **3.1 Effect of Different Salt Concentrations on** *Camelina* **Seedlings**

Seven-day-old *Camelina* seedlings (green cotyledons and 2-4 mm of root) were put in petri dishes with different NaCl concentrations (0, 50, 75, 100, 125 and 150 mM). After 3 day of stress, the appearance of seedlings was checked (Fig. 1). At higher concentrations (100-150 mM),

the seedlings were flabby, no root growth was observed and showed yellowish leaves. At 50 mM of salt, the seedlings appeared very similar to control without NaCl while, at 75 mM, they showed the first sign of cotyledons bleaching and no root growth was observed and, in general, the tissues seemed less firm.

The results indicate that 75 mM is a critical concentration of salt for seedling development in *Camelina sativa*. At this concentration, the effect observed did not probably depend on salt concentration in the growing tissues, but rather to a response to the osmolarity of the external solution [19]. Bleaching effect on cotyledons probably depends on the rate of accumulation of Na<sup>+</sup>, and on the effectiveness of Na<sup>+</sup> compartmentation within cotyledon cells [20].

# **3.2 Levels of Soluble Sugars and Total Amino Acids at Different Salt Concentrations**

The levels of soluble sugars and total amino acids in the presence of different concentrations of NaCl (0, 50, 75, 100, 125 and 150 mM) are shown in Fig. 2. Both the content of sugars and amino acids were enhanced with increasing salt concentration up to 75 mM. At higher concentrations of NaCl, their levels begin to drop, and this decrease turned out stronger on soluble sugars. While the content of amino acids resulted lower than the control (no salt) only at 150 mM NaCl, the level of soluble sugars already at 100 mM NaCl was lower than the control, continuing to fall with increasing salt concentration. The results indicate that *Camelina* seedlings are able to well counteract the increase in osmotic pressure due to the presence of salt up to 75 mM and then manifest problems at higher concentrations, starting likely to excessively consume sugars.

Sugars contribute up to 50% of the total osmotic potential in glycophytes subject to saline conditions [21]. *Camelina* showed that, above 75 mM NaCl, is unable to maintain the sugar content (Fig. 2) even if the level of total amino acids was still high (probably for protein synthesis inhibition). Therefore, as generally observed in glycophytes [22], *Camelina* was unable to tolerate salt stress.

# **3.3 Polyamine Levels in the Presence of Different Salt Concentrations**

PAs have been deemed important in preparing the plant for stress tolerance and to directly aid in ameliorating the causes of stress [23]. In Fig. 3, the level of total PAs in the presence of different salt concentrations is shown. Total PAs increased at 50 mM NaCl but then decreased, resulting below the control without salt already at 100 mM.



**Fig. 1. Effect of 3 day treatment with different NaCl concentrations (0, 50, 75, 100, 125 and 150 mM) on** *Camelina* **seedlings**



**Fig. 2. Effect of different NaCl concentrations on soluble sugars (open bars) and total amino acids (solid bars). Data are expressed on a fresh weight (FW) basis**



**Fig. 3. Effect of different NaCl concentrations on total PA level. Data are expressed on a fresh weight (FW) basis**

The PA increase at 50 mM was mainly due to the increase in Spm level (Fig. 4), while all the PA species decreased from 75 mM forward (the decrease was more pronounced on Spd and Spm levels). This causes a decline in the polyamine/diamine ratio from 3.66 at 75 mM NaCl up to 2.98 at 150 mM NaCl.

Several studies have emphasized the<br>contribution of PAs in tolerance and/or contribution of PAs in

amelioration of stress in plants [24-28]. In particular, in *Arabidopsis* the PA Spm protected against salinity and drought stress [13,14]. Moreover, when Spm was exogenously applied, it restored a normal germination of seeds of sunflower and sorghum in the presence of NaCl [29,30]. Here, we showed that *Camelina* seedlings increased the level of Spm only at 50 mM NaCl, but then they were unable to maintain PA levels at higher salt concentrations

(Figs. 3-4). Therefore, the decline of PAs goes hand in hand with the rise of intolerance to salt stress of *Camelina* seedlings as evidenced in Fig. 1.

#### **3.4 Effect of Exogenous Spm on Salinity Sensitivity of** *Camelina* **Seedlings**

To test the importance of PAs in the tolerance of salinity, 7-day-old seedlings *Camelina* were subjected to different NaCl concentrations (0, 75 and 125 mM) in the presence or absence of 1 mM Spm (Fig. 5). The presence of exogenous Spm did not affect the control seedllings while those with 75 mM NaCl were normal and indistinguishable from the control without salt (those without Spm appeared suffering). A 125 mM NaCl, the presence of Spm kept the seedlings more green than those without Spm which were completely decoloured. The results further confirm the role of PAs and, in particular, of SPM in salt stress tolerance.







**Fig. 5. Effect of 3-day treatment with different NaCl concentrations (0, 75 and 125 mM) of**  *Camelina* **seedlings in the presence or absence of Spm (1 mM)**

# **4. CONCLUSION**

*Camelina sativa* is a glycophyte sensitive to salt (NaCl) at 75 mM. At this concentration, *Camelina* is still able to counteract the osmotic pressure due to salt but begins to show damages to seedlings after 3 days of treatment. The content of PAs starts to decrease along with the appearance of damage to seedlings. In particular, the content of Spm definitely increases at 50 mM when the seedlings appear beautiful and then decreases along with the other PAs. The administration of exogenous Spm during the salt stress improved tolerance in *Camelina* seedlings. These data further support the role of Spm protection against salinity.

# **ACKNOWLEDGEMENTS**

The authors are gratefully to Regione Lombardia (Italy), agreement Regione/CNR, project 2: 'Risorse biologiche e tecnologie innovative per lo sviluppo sostenibile del sistema agro-alimentare' for supporting the work.

# **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

# **REFERENCES**

1. Bupta B, Huang B. Mechanism of salinity tolerance in plants: Physiological, biochemical, and molecular characterization. Int J Genomics. 2014; 1-18.

Available:http://dx.doi.org/10.1155/2014/70 1596

- 2. Munns R. Genes and salt tolerance: Bringing them together. New Phytologist. 2005;167(3):645–663.
- 3. Rozema J, Flowers T. Ecology: Crops for a salinized world. Science. 2008; 322(5907):1478–1480.
- 4. Reggiani R, Bozo S, Bertani A. The effect of salinity on early seedling growth of seeds of three wheat (*Triticum aestivum* L.) cultivars. Can J Plant Sci. 1995;75: 175-177.
- 5. Matysik J, Alia A, Bhalu B, Mohanty P. Molecular mechanisms of quenching of reactive oxygen species by proline under stress in plants. Curr Sci. 2002;82(5): 525–532.
- 6. Saxena SC, Kaur H, Verma P, Bhanu P, Petla BP, Andugula VR, Majee M. Osmoprotectants: Potential for crop improvement under adverse conditions. In: Tuteja N, Gill SS, editors. Plant Acclimation to Environmental Stress, New York: Springer. 2013;197–232.
- 7. Parida AK, Das AB, Mohanty P. Investigations on the antioxidative defence responses to NaCl stress in a mangrove,<br>Bruguiera parviflora: Differential *Bruguiera parviflora:*<br>
regulations of isoforms regulations of isoforms of some antioxidative enzymes. Plant Growth Regul. 2004;42(3):213–226.
- 8. Reggiani R, Aurisano N, Mattana M, Bertani A. Influence of  $K^+$  ions on polyamine level in wheat seedlings. J Plant Physiol. 1993;141:136-140.
- 9. Begum F, Karmoker JL, Fattah QA, Maniruzzaman AFM. The effect of salinity on germination and its correlation with  $K^f$ , Na<sup>+</sup>, CI<sup>-</sup> accumulation in germinating seeds of *Triticum aestivum* L. cv. Akbar. Plant Cell Physiol. 1992;33:1009-1014.
- 10. Reggiani R, Bozo S, Bertani A. Changes in polyamine metabolism in seedling of three wheat (*Triticum aestivum* L.) cultivars differing in salt sensitivity. Plant Sci. 1994;102:121-126.
- 11. Bouchereau A, Aziz A, Larher F, Martin-Tanguy J. Polyamines and environmental challenges: Recent development. Plant Sci. 1999;140:103–125.
- 12. Cohen SS. A guide to the polyamines. New York: Oxford University Press; 1998.
- 13. Yamaguchi K, Takahashi Y, Berberich T, Imaic A, Miyazakia A, Takahashic T, Michaeld A, Kusano T. The polyamine spermine protects against high salt stress in Arabidopsis thaliana. FEBS Lett. 2006; 580(30):6783–6788.
- 14. Kusano T, Yamaguchi K, Berberich T, Takahashi Y. The polyamine spermine rescues Arabidopsis from salinity and drought stresses. Plant Signaling Behavior. 2007;2(4):251–252.
- 15. Moore S. Amino acid analysis: Aqueous dimethyl sulfoxide as solvent for ninhydrin reaction. J Biol Chem. 1968;243: 6281-6283.
- 16. Aurisano N, Bertani A, Reggiani R. Involvement of calcium and calmodulin in protein and amino acid metabolism in rice roots under anoxia. Plant Cell Physiol. 1995;36(8):1525-1529.
- 17. Hansen J, Møller IB. Percolation of starch and soluble carbohydrates from plant

tissue for quantitative determination with anthrone. Anal Biochem. 1975;68:87-94.

- 18. Reggiani R, Hochkoeppler A, Bertani A. Polyamines in rice seedlings under oxygen-deficit stress. Plant Physiol. 1989; 91:1197-1201.
- 19. Munns R. Comparative physiology of salt and water stress. Plant Cell Environ. 2002; 25:239–250.
- 20. Parvaiz A, Satyawati S. Salt stress and phyto-biochemical responses – A review. Plant Soil Environ. 2008;54(3):89–99.
- 21. Cram WJ. Negative feedback regulation of transport in cells. The maintenance of turgor, volume and nutrient supply. In: Luttge U, Pitman MG, editors. Encyclopaedia of Plant Physiology, New Series, Vol. 2. Berlin: Springer-Verlag; 1976.
- 22. Sairam RK, Tyagi A. Physiology and molecular biology of salinity stress tolerance in plants. Curr Sci. 2004;86: 407–421.
- 23. Minocha R, Majumdar R, Minocha SC. Polyamines and abiotic stress in plants: A complex relationship. Frontiers Plant Sci. 2014;5:1-17.
- 24. Alcázar R, Marco F, Cuevas JC, Patron M, Ferrando A, Carrasco P, Tiburcio AF, Altabella T. Involvement of polyamines in plant response to abiotic stress. Biotechnol Lett. 2006;28,1867–1876.

Available:http://dx.doi.org/10.1007/s10529- 006-9179-3

25. Takahashi T, Kakehi J. Polyamines: Ubiquitous polycations with unique roles in growth and stress responses. Ann Bot. 2010;105:1–6.

Available:http://dx.doi.org/10.1093/aob/mc p259

- 26. Hussain SS, Ali M, Ahmad M, Siddique KHM. Polyamines: Natural and engineered abiotic and biotic stress tolerance in plants. Biotechnol Adv. 2011;29:300–311. Available:http://dx.doi.org/10.1016/j.biotec hadv.2011.01.003
- 27. Gupta K, Dey A, Gupta B. Plant polyamines in abiotic stress responses. Acta Physiol Plant. 2013;35:2015–2036. Available:http://dx.doi.org/10.1007/s11738- 013-1239-4
- 28. Shi H, Chan Z. Improvement of plant abiotic stress tolerance through modulation of the polyamine pathway. J Integr Plant Biol. 2014;56:114–121. Available:http://dx.doi.org/10.1111/jipb.121 28
- 29. Mutlu F, Bozcuk S. Effects of exogenous spermine on seed germination and early growth of sunflower seeds under salinity stress. Turk J Biol. 2000;24:635-643.
- 30. Chai YY, Jiang CD, Shi L, Shi TS, Gu WB. Effects of exogenous spermine on sweet sorghum during germination under salinity. Biol Plant. 2010;54(1):145-148.

 $\_$  , and the set of th *© 2015 Russo and Reggiani; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.*

> *Peer-review history: The peer review history for this paper can be accessed here: http://sciencedomain.org/review-history/10695*