

Susceptibility of some Cruciferae crops cultivars against the root-knot nematode, *Meloidogyne Javanica*

Abdel-Hafeez A. R. *, Mahmoud N. A., El-Mesalamy A. F.

Agricultural Zoology and Nematology Department, Faculty of Agriculture, Al-Azhar University, Assiut 71524, Egypt

Abstract

Ten Cruciferae crops species were evaluated for their host's response to root-knot nematode, *Meloidogyne javanica*. The nematode succeeded in developing and multiplying on all most tested cultivars (cvs.). The ten plant species were divided by their degree of susceptibility to root-knot nematode, *Meloidogyne javanica* as follows: Radish Baladi cv., Cabbage Brunswick cv. and Cabbage Japanese cv. were very resistant host to nematode infection. The calculated values of the numbers of galls per root and rates of nematode reproduction on such cultivars were (24.50–0.15), (9.00–0.04) and (6.25–0.02) respectively. While Cabbage Sabeini cv., Cauliflower Sultani cv. and Cabbage Dutch cv. were slightly resistant hosts to nematode infection. The calculated values of the numbers of galls per root and rates of nematode reproduction on such cultivars recorded moderate values were 63.75–0.58, 62.25–0.02, and 61.75–0.76 respectively. Turnip Baladi cv., Turnip Japanese cv., Red Radish cv. and Cauliflower Chinese cv. were susceptible hosts to nematode infection, their calculated values of the numbers of galls per root and rates of nematode reproduction on such cultivars were 290.75–9.29, 232.75–4.92, 169.50–4.29 and 120.75–0.29, respectively.

Keywords: *Meloidogyne javanica*, susceptibility, Cruciferae, cultivar.

*Corresponding author: Abdel-Hafeez A. R.,
E-mail address: abdelhafeez@azhar.edu.eg

1. Introduction

Plant-parasitic nematodes, especially root-knot nematodes (*Meloidogyne*), are among the most important soil-borne pests that cause severe yield and/or quality losses to a wide range of agricultural crops (Jones et al., 2013). Moreover, more than 2000 species of plants have already been identified as hosts for root-knot nematodes (Bird et al., 2008), and more than 90 species of root-knot nematodes are described so far (Hunt and Handoo, 2009). Ten of these species are of importance from an agricultural point of view, and four species, *Meloidogyne hapla*, *M. arenaria*, *M. javanica* and *M. incognita*, are considered as major pests in many areas of the world (EPPO/OEPP, 2004). Root-knot nematodes cause considerable problems in intensive agricultural cropping (Caillaud et al., 2008) leading to substantial economic losses, mainly as a result of quality damage, to tuber-forming annual crops (Bird et al., 2008; Boydston et al., 2007). Interestingly, cabbage (*Brassica oleracea* var. *capitata*) is that is rich in vitamin C, it is regarded as the most significant member of the *Brassicaceae* (Cruciferae) family and has remained one of the world's leading vegetable crops. Also, cabbage is an economically important vegetable crop in several Mediterranean countries (FAOSTAT, 2010). In the Mediterranean basin, under field conditions, cabbage is usually transplanted in late summer and harvested in early spring, and is exposed to infection by several diseases, particularly those caused by soil-borne pathogens. Several diseases have been reported damaging cabbage worldwide. While some of them

may simply cause minor spotting, others can be devastating for the crop. Among them plant-parasitic nematodes, particularly root-knot nematodes, *Meloidogyne* spp., are considered major diseases (Rimmer et al., 2007). *Meloidogyne* spp. prevailing in the Mediterranean basin, such as *M. arenaria*, *M. incognita* and *M. javanica* are warm climate species. However, the low soil temperatures reached in the Mediterranean basin in fall and winter are not suitable for infection and development of these *Meloidogyne* species (Moens et al., 2009) and, therefore, the incidence of root-knot nematodes in cabbage in this region is usually low (Buczacki et al., 1978). The pathogenicity of the sugar beet cyst nematode *Heterodera schachtii* and the root-knot nematodes *Meloidogyne arenaria*, *M. incognita* and *M. javanica* on cabbage cvs. Balady, Brunswick and Ganzouri, cauliflower cv. Balady, turnip cv. Balady, and radish cv. Balady was determined in several greenhouse tests. The results showed that the tested cruciferous plant cultivars were either susceptible or highly susceptible to the tested nematodes except radish cv. balady, which was moderately resistant to *H. schachtii* and moderately susceptible to the tested root-knot nematode species (Ibrahim et al., 2013). Although some studies exist on the host status of various Brassicaceae to root-knot nematodes (Khan and Khan, 1991; McSorley and Frederick, 1995; Netscher and Sikora, 1990), little information exists on the host status of Egyptian crucifers to *Meloidogyne* spp., or control measures for them (Ibrahim et al., 2013). Other vegetables are reported in the literature as

susceptible to root-knot nematodes, including brassicaceae, solanaceae, cucurbitaceae and liliaceae (Brito *et al.*, 2007; McSorley and Frederick, 1995, Ponte *et al.*, 1996; Walker, 2002). The root-knot nematode species most damaging to cabbage are *M. arenaria* (Neal) Chitw., *M. artiellia* Franklin, *M. hapla* Chitw., *M. incognita* (Kofoid ET White) Chitw., *M. javanica* (Treub) Chitw. and *M. enterolobii* Yan ET Eisenback (= *M. mayaguensis* Rammah ET Hirschmann) (Abrantes *et al.*, 1994; Potter and Olthof, 1993; Sikora and Fernández, 2005). *Meloidogyne spp.* prevailing in the Mediterranean basin, such as *M. arenaria*, *M. incognita* and *M. javanica* are warm climate species. However, the low soil temperatures reached in the Mediterranean basin in fall and winter are not suitable for infection and development of these *Meloidogyne species* (Moens *et al.*, 2009). The results showed that *M. incognita* and *M. javanica* responded in a similar fashion to the different cover cultivars. Indian mustard (*Brassica juncea*) and turnip (*B. rapa*) were generally good hosts, whereas most oil radish cultivars (*Raphanus sativus ssp. oleiferus*) were poor hosts. However, some oil radish cultivars were among the best hosts for *M. hapla*. The arugula (*Eruca sativa*) cultivar Nemat was a poor host for all three nematode species tested (Edwards and Ploeg, 2014). Egg masses from the Brassicaceae species contained fewer eggs than tomato egg masses. Differences were not significant at 5 weeks, but all Brassicaceae species differed from tomato at 6 weeks. Only rape cv. Korina differed significantly from tomato at 7 weeks and hybrid turnip cv. Purple Top and tomato did not differ

significantly at 8 weeks (McLeod *et al.*, 2001). Although Brassicaceae crops are hosts, they are poorly invaded and suppress nematode growth and development, thereby reducing the risk of *M. javanica* increasing on Brassicaceae crops (Stanton and Eyres, 1994).

2. Materials and methods

2.1 Culture of nematodes

Root knot nematode *Meloidogyne javanica* was selected as test organism. The culture of root knot nematode, *Meloidogyne javanica*, is maintained on potted eggplant plants in the glasshouse. The infected plants will be uprooted, carefully washed in running tap water and egg- masses will be collected into Petri dishes containing distilled water.

2.2 Greenhouse screening tests

Seeds of ten Cruciferae crops Sultani and Chinese, (Cauliflower), Sabeini, Dutch, Japanese and Brunswick (Cabbage), Red and Baladi (Radish) and Baladi and Japanese (Turnip) were germinated in 15 cm diameter clay pots filled with a 2:1 mixture of loamy sand soil. Six weeks - old, germinated seedlings were thinned to one healthy plant per pot. Each pot was inoculated with approximately 3000 newly hatched juveniles of each *Meloidogyne javanica* per plant by pipetting the nematode suspension in three holes around the root system. Four pots were used as replicates for each plant species. Inocula of each nematode species

were obtained from available pure stock cultures maintained on suitable hosts in a greenhouse. Each plant species was replicated four times including those kept non-infected serving as check. All pots were then, arranged in a randomized block design on a bench in a greenhouse. All plants were grown during the normal growing seasons at greenhouse temperature of 20 ± 5 °C. Fifty days after inoculation, all plants were harvested, and the root system of each plant was carefully removed gently washed in water and stained in lactophenol acid fuchsin (Goodey, 1957). The number of juveniles in soil per pot, galls, nematode developmental stages on root, egg-masses per root was counted. Eggs of ten randomly selected egg-masses of each root system were also counted by sodium hypochlorite. The rate of nematode reproduction was calculated. Plant growth criteria involving length and fresh weight of both roots and shoots were calculated. The percentages of reduction of such parameters of each plant were also collected. Root gall index values were estimated according to the following scales: 0 = 0 galls, 1 = 1–2 galls, 2 = 3–10 galls, 3 = 11–30 galls, 4 = 31–100 galls and 5 = > 100 galls (Taylor and Sasser, 1978). The host category of the tested crop cultivars plants infected with the root-knot nematode, based on root galls index ranges was determined according to Hadisoeganda and Sasser (1982), as follow 0 = Immune host (I), 0.0–1.0 = highly resistant (HR), 1.1–3.0 = very resistant (VR), 3.1–3.5 = moderately

resistant (MR), 3.6–4.0 = slightly resistant (SR), 4.1–5.0 = susceptible (S).

3. Results and Discussion

Ten Cruciferae crops species were exposed under greenhouse conditions against attack by the root-knot nematode *Meloidogyne javanica* it is evident that, the nematode reproduction and nematode fecundity were greatly affected by host type. Results in Table (1) showed that *M. javanica* larvae succeeded in developing and multiplying on all the tested Cruciferae family cultivars. Therefore, the nematode population was significantly affected by the tested cultivars under the same conditions, thereupon Turnip Baladi cv., Turnip Japanese cv., Red Radish cv. and Cauliflower Chinese cv. gained the highest values of number of galls per root, nematode juveniles in soil, nematode developmental stages per root, egg-masses per root, eggs per egg-mass, nematode final population, and rate of nematode reproduction among at the tested cultivars. Significant differences ($P \geq 0.05$) in the nematode criteria on such plant cultivars were observed when compared with those of the other tested cultivars. Also, their calculated values of the numbers of galls per root and rates of nematode reproduction on such cultivars were 290.75–9.29, 232.75–4.92, 169.50–4.29 and 120.75–0.29, respectively. While, tested Cabbage Sabeini, Cauliflower Sultani and Cabbage Dutch cultivars the recorded moderate values of

the numbers of galls per root and rates of nematode reproduction on such cultivars were 63.75–0.58, 62.25–0.02, and 61.75–0.76, respectively. On the other hand, Radish Baladi, Cabbage Brunswick and Cabbage Japanese cultivars supported the lowest values of nematode juveniles in soil, number of galls per root, nematode

developmental stages per root, egg-masses per root, eggs per egg-mass, rate of nematode reproduction and nematode final population. The calculated values of the numbers of galls per root and rates of nematode reproduction on such cultivars were 24.50–0.15, 9.00–0.04 and 6.25–0.02, respectively (Table 2).

Table (1): Susceptibility of some Cruciferae crop cultivars to the root-knot nematode *Meloidogyne javanica* under greenhouse conditions.

Scientific name	Common name	Cultivars	Number of galls/root	Nematode population						Nematode final population (P _i)	Rate of nematode reproduction (P _r /P _i)	Egg production (%)	Host category
				Juveniles in soil/pot	Nematode develop stage/root	Adult female/root	Number of egg masses /root	Number of eggs/egg mass					
<i>Brassica oleracea</i> var. <i>botrytis</i>	Cauliflower	Sultani	62.25 e	0.00 e	56.50 cd	7.00 e	5.00 e	0.00 f	63.50	0.02	0.00	SR	
		Chinese	120.75 d	60.00 de	88.75 ab	30.00 c	27.75 c	25.25 f	879.44	0.29	2.61	S	
<i>Brassica oleracea</i> var. <i>capitata</i>	Cabbage	Sabeini	63.75 e	65.50 de	29.00 def	31.00 c	24.75 c	65.00 de	1734.25	0.58	5.99	SR	
		Dutch	61.75 e	117.25 d	39.00 cde	24.50 c	25.75 c	81.75 cd	2285.81	0.76	7.84	SR	
		Japanese	6.25 f	0.00 e	5.75 f	2.00 e	1.50 e	33.75 ef	58.38	0.02	0.19	VR	
		Brunswick	9.00 f	0.00 e	6.00 f	4.75 c	2.50 c	43.00 ef	118.25	0.04	0.40	VR	
<i>Raphanus sativus</i>	Radish	Red Radish	169.50 c	617.50 b	62.00 bc	106.50 b	83.25 b	145.25 b	12878.06	4.29	45.04	S	
		Baladi	24.50 ef	266.50 c	13.25 ef	9.00 c	6.00 c	26.25 f	446.25	0.15	0.59	VR	
<i>Brassica rapa</i> var. <i>rapifera</i>	Turnip	Baladi	290.75 a	736.50 a	108.00 a	182.00 a	148.75 a	180.50 a	27875.88	9.29	100.00	S	
		Japanese	232.75 b	652.50 b	90.25 ab	152.75 a	135.50 a	102.25 c	14750.38	4.92	51.60	S	

Means in each column followed by the same letter are not significantly different by (p=0.05) according to Duncan's multiple range test.

Table (2): Plant growth of some Cruciferae crop cultivars as influenced with the infection of the root-knot nematode *Meloidogyne javanica* under greenhouse conditions.

Scientific name	Common name	Cultivars	Length in (cm)									Fresh weight in (gm)						Host Category
			Shoot			Root			Shoot			Root						
			Infected	Non-infected	Decr. %	Infected	Non-infected	Decr. %	Infected	Non-infected	Decr. %	Infected	Non-infected	Decr. %				
<i>Brassica oleracea</i> var. <i>botrytis</i>	Cauliflower	Sultani	29.75	30.75	3.25	27.11	27.00	-0.41	15.33	16.31	6.36	3.50	3.81	8.86	SR			
		Chinese	19.50	20.00	2.50	24.75	26.75	7.48	12.41	12.40	-0.10	2.62	2.65	1.15	S			
		Sabeini	19.25	21.00	8.33	9.00	9.25	2.70	5.09	5.99	17.57	0.78*	1.03	32.80	SR			
<i>Brassica oleracea</i> var. <i>capitata</i>	Cabbage	Dutch	19.75*	23.25	15.05	18.50	19.00	2.63	11.36	11.51	1.37	6.33	7.56	19.52	SR			
		Japanese	23.75	24.25	2.06	16.50	16.50	0.00	14.71	14.49	-1.50	3.74	4.66	24.52	VR			
		Brunswick	23.25*	24.25	4.12	11.00	11.50	4.35	12.38	13.39	8.10	2.61	2.62	0.48	VR			
<i>Raphanus sativus</i>	Radish	Red Radish	24.00*	31.25	23.20	29.25*	33.00	11.36	9.65*	13.27	37.46	3.43	5.00	45.73	S			
		Baladi	25.75	27.75	7.21	25.25	27.00	6.48	10.77	13.85	28.55	2.74	3.01	9.85	VR			
<i>Brassica rapa</i> var. <i>rapifera</i>	Turnip	Baladi	25.00	32.25	22.48	22.50**	30.25	25.62	9.76	11.07	13.40	13.47	14.43	7.13	S			
		Japanese	20.25*	26.50	23.58	21.00*	24.50	14.29	7.42	10.55	42.18	2.75	3.35	21.73	S			

* Significant at 0.05 level of probability. ** Highly significant at 0.01 level of probability.

These results are in accordance with findings of McLeod *et al.* (2001), McSorley and Frederick (1995) and Ibrahim *et al.* (2013). The ten plant species were divided by their degree of susceptibility to *Meloidogyne javanica* as follows:

- Radish Baladi, Cabbage Brunswick and Cabbage Japanese cultivars were

very resistant host to nematode infection.

- Cabbage Sabeini, Cauliflower Sultani and Cabbage Dutch were slightly resistant host to nematode infection.
- Turnip Japanese, Red Radish and Cauliflower Chinese cultivars were susceptible host to nematode infection.

References

- Abd Elgalil, M. A. S. and Abdel-Gawad, A. M. A. (2020), "Irrigation water regime and manure extract for wheat production grown under drip irrigation", *Archives of Agriculture Sciences Journal*, Vol. 3 No. 3, pp. 306–318.
- Abrantes, I. M., Vovlas N., de Santos M. S. N. A. and Espirito Santo, S. N. (1994), "*Meloidogyne arenaria* and *Plasmiodiophora brassicae*, causal agents of gall development on cabbage roots in the Republica Democrática de Sao Tomé e Príncipe", *Nematologia Mediterranea*, Vol. 22, pp. 115–118.
- Bird, D. M., Opperman, C. H. and Williamson, V. M. (2008), "Plant infection by root-knot nematode", *Plant Cell Monographs*, Vol. 10, pp. 1–5.
- Boydston, R. A., Mojtahedi, H., Brown, C. R., Anderson, T. and Riga, E. (2007), "Hairy nightshade undermines resistance of potato breeding lines to Columbia root-knot nematode", *American Journal of Potato Research*, Vol. 84, pp. 245–251.
- Brito, J. A., Stanley J. D., Mendes M. L., Cetintas R. and Dickson D. W. (2007), "Host status of selected cultivated plants to *Meloidogyne mayaguensis* in Florida", *Nematropica*, Vol. 37, pp. 65–71.
- Buczacki, S. T., Ockendon, J. G. and Freeman, G. H. (1978), "An analysis of some effects of light and soil temperature on clubroot disease", *Annals of Applied Biology*, Vol. 88, pp. 229–238.
- Caillaud, M. C., Dubreuil, G., Quentin, M., Perfus-Barbeoch, L., Lecomte, P., de Almeida Engler, J., Abad, P., Rosso, M. N. and Favery, B. (2008), "Root-knot nematodes manipulate plant cell function during a compatible interaction", *Journal of Plant Physiology*, Vol. 165, pp. 104–113.
- Edwards, S. and Ploeg, A. (2014), "Evaluation of 31 potential biofumigant brassicaceous plants as hosts for three *Meloidogyne species*", *Journal of Nematology*, Vol. 46 No. 3, pp. 287–295.
- EPPO (2004), "Diagnostic protocols for regulated pests, European and Mediterranean Plant Protection Organization", *EPPO Bulletin*, Vol. 34, pp. 315–320.
- FAOSTAT (2010), *Faostat production statistics of crops (FAOSTAT)*, Food and Agriculture Organization of the United Nations, Available at <http://faostat.fao.org>.
- Goodey, J. B. (1957), "*Hoplolaimus proporicus* n. sp., (Hoplolaiminae: Tylenchida)", *Nematologica*, Vol. 2, pp. 108–113.
- Hadisoeganda, W. W. and Sasser, J. N.

- (1982), "Resistance of tomato, bean, southern pea, and garden pea cultivars to root knot nematodes based on host suitability", *Plant Disease*, Vol. 66 No. 2, pp. 145–150.
- Hunt, D. J. and Handoo, Z. A. (2009), "Taxonomy, identification and principal species", In: Perry, R. N., Moens, M. and Starr, J. L. (Eds). *Root-knot nematodes*, CAB International, Wallingford, UK, pp. 55–97.
- Ibrahim, I. K. A., Basyony, A. B. A., Handoo, Z. A. and Chitwood, D. J. (2013), "Pathogenicity and control of *Heterodera schachtii* and *Meloidogyne* spp. on some cruciferous plant cultivars", *International Journal of Nematology*, Vol. 23 No. 1, pp. 73–81.
- Jones, J. T., Haegeman, A., Danchin, E. G. J., Gaur, H. S., Helder, J., Jones, M. G. K., Kikuchi, T., López, R. M., Rius, J. E. P., Wesemael, W. M. L. and Perry, R. N. (2013), "Top 10 plant-parasitic nematodes in molecular plant pathology", Vol. 14 No. 9, pp. 946–961.
- Khan, A. A. and Khan, M. W. (1991), "Reaction of cauliflower cultivars to *Meloidogyne javanica* and races of *Meloidogyne incognita*", *Nematropica*, Vol. 21, pp. 161–166.
- McLeod, R. W., Kirkegaard, J. A. and Steel, C. C. (2001), "Invasion, development, growth and egg laying by *Meloidogyne javanica* in Brassicaceae crops", *Nematology*, Vol. 3 No. 5, pp. 463–472.
- Mcsorley, R. and Frederick, J. J. (1995), "Responses of some common Cruciferae to root-knot nematodes", *Journal of Nematology*, Vol. 27, pp. 550–554.
- Moens, M., Perry, R. N. and Starr, J. L. (2009), "*Meloidogyne* species – a diverse group of novel and important plant parasites", In: *Root-Knot Nematodes*, Moens M., Perry R.N. and Starr J.L. eds., CABI Publishing, Wallingford, UK, pp. 1–17.
- Netscher, C. and Sikora, R. A. (1990), "Nematode parasites of vegetables", In: *Plant Parasitic Nematodes in Subtropical and Tropical Agriculture*, Luc, M., Sikora, R. A. and Bridge J. eds., CAB International, Wallingford, UK, pp. 237–283.
- Ponte, J. J., Holanda, Y. C. A. and Aragão, M. L. (1996), "Adendo ao catálogo de plantas hospedeiras de *Meloidogyne* no Brasil", *Nematologia Brasileira* 20: 73–81.
- Potter, J. W. and Olthof, T. H. A. (1993), "Nematode pest of vegetable crops", In: *Plant Parasitic Nematodes in Temperate Agriculture*, Evans K., Trudgill D.L. and Webster J.M. eds., CAB International, Wallingford, UK, pp. 171–207.
- Rimmer, S. R., Shattuck, V. I. and

- Buchwaldt, L. (2007), *Compendium of Brassica Diseases*, The American Phytopathological Society, St. Paul, MN, USA, pp. 117.
- Sikora, R. A. and Fernández, E. (2005), "Nematodes parasites of vegetables", *In: Plant Parasitic Nematodes in Subtropical and Tropical Agriculture*, Luc M., Sikora R.A. and Bridge J. eds., CABI Publishing, Wallingford, UK, pp. 319–392
- Stanton, J. M. and Eyre, S. M. (1994), "Hatching of Western Australian populations of cereal cyst nematode, *Heterodera avenae*, and effects of sowing time and method of sowing on yield of wheat", *Australasian Plant Pathology*, Vol. 23, pp. 1–7.
- Taylor, A. L. and Sasser, J. N. (1978), *Biology, identification, and control of root-knot nematodes (Meloidogyne spp.)*, North Carolina State University Graphics, Raleigh, USA
- Walker, J. T. (2002), "Susceptibility of eight herbs to common root-knot nematodes", *Journal of Environmental Horticulture*, Vol. 20, pp. 101–110.